

E.ADEM v0.0.4.4

2009.7.18

2009.5.3

75 days

2009.7.18

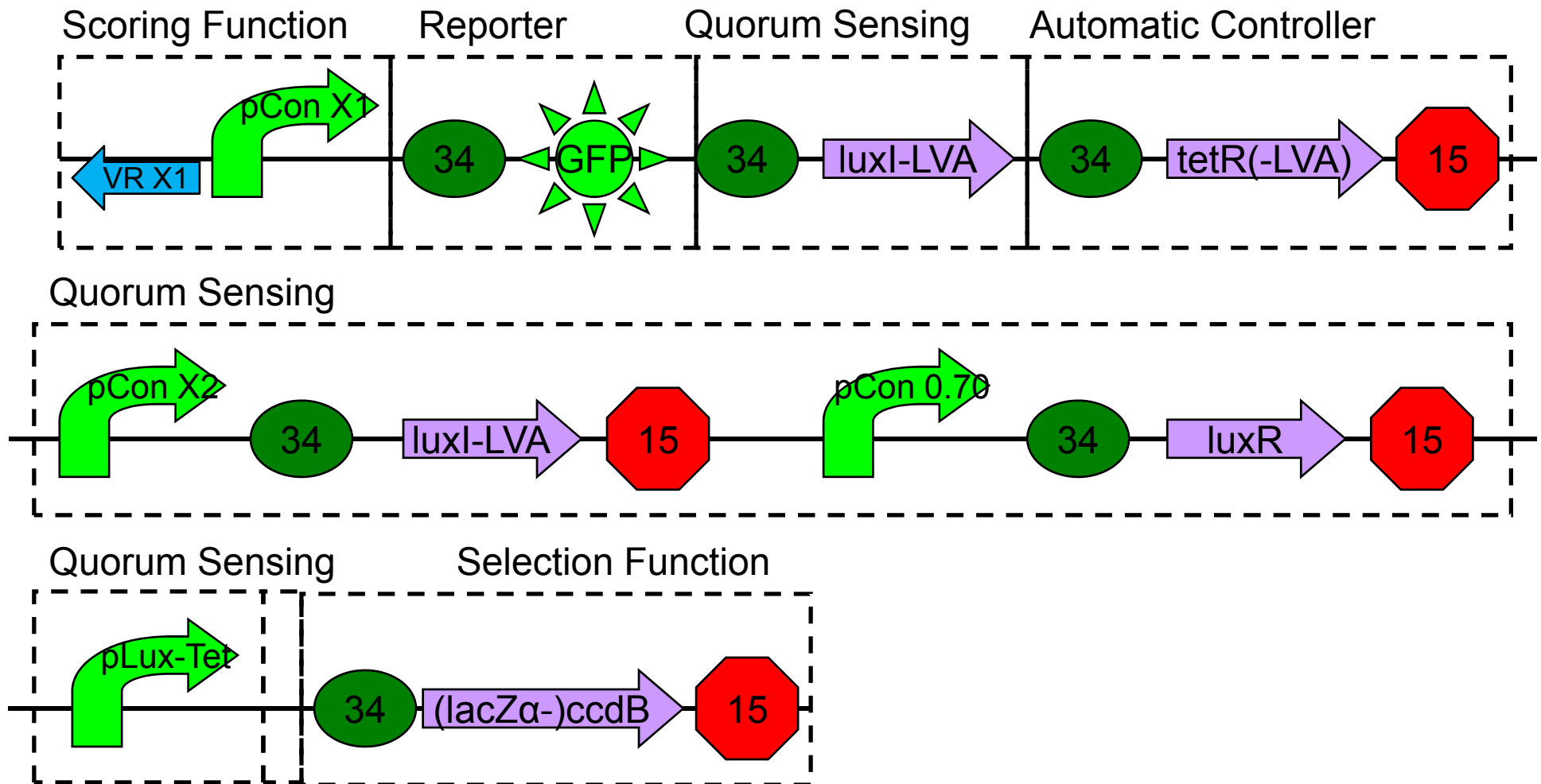
103 days

2009.10.30

Outline

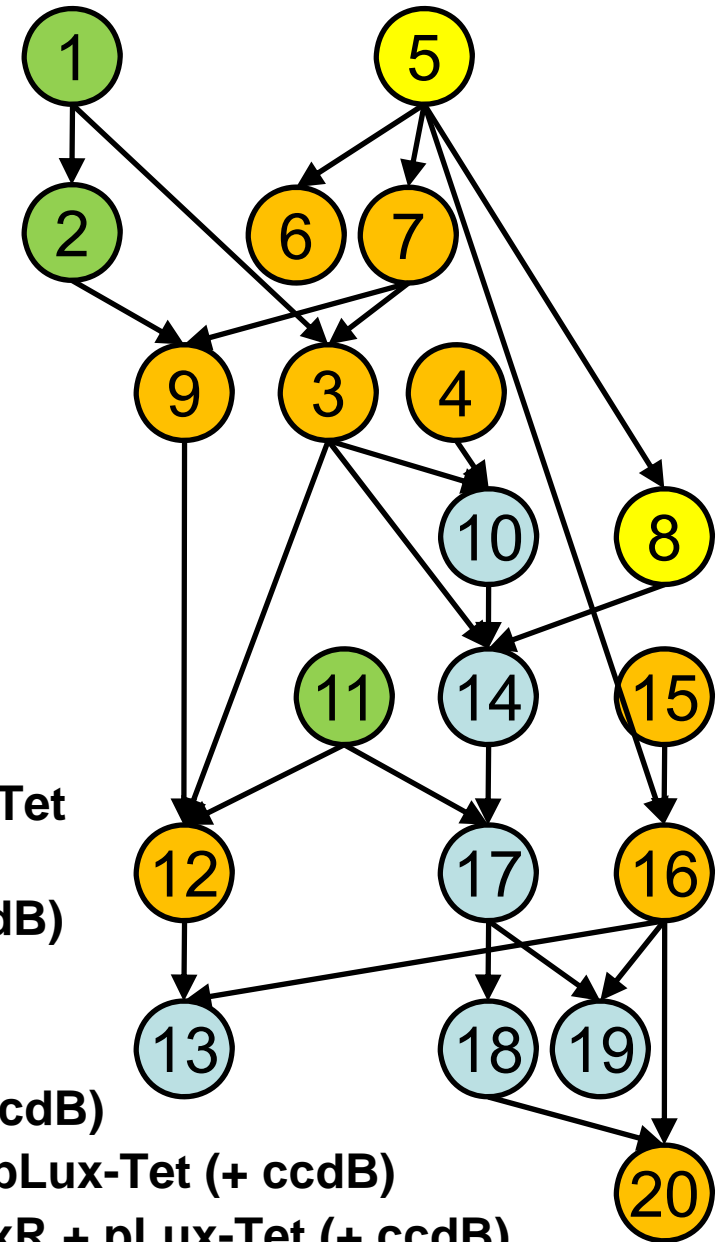
- Assembly
- Measurement
 - General Conditions
 - GFP
 - AHL
 - CcdB
 - LacZ α
- Wiki
 - Team project description
 - Notebook
- Instructional Videos

Assembly

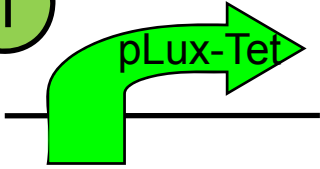


1. pLux-Tet
2. pLux-Tet + GFP
3. pCon + luxR + pLux-Tet
4. ccdB × 2
5. pCon × 8
6. pCon × 8 + GFP
7. pCon + luxR
8. pCon × 7 + luxI (AHL detection by 9 or GC-MS ?)
9. pCon + luxR + pLux-Tet + GFP (AHL)
10. pCon + luxR + pLux-Tet + ccdB × 2 (AHL)
11. tetR × 2
12. tetR × 2 + pCon + luxR + pLux-Tet (+ GFP)
13. (VR + pCon) × 8 + tetR × 2 + pCon + luxR + pLux-Tet (+ GFP) (AHL/aTc)
14. (pCon × 7 +) luxI + pCon + luxR + pLux-Tet (+ ccdB)
15. VR × 8
16. (VR + pCon) × 8
17. tetR + pCon + luxI + pCon + luxR + pLux-Tet (+ ccdB)
18. GFP + luxI + tetR + pCon + luxI + pCon + luxR + pLux-Tet (+ ccdB)
19. (VR + pCon) × 8 + tetR + pCon + luxI + pCon + luxR + pLux-Tet (+ ccdB)
20. (VR + pCon) × 8 + GFP + luxI + tetR + pCon + luxI + pCon + luxR + pLux-Tet (+ ccdB)

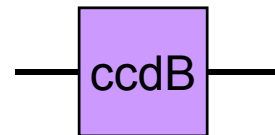
Waiting
Working
Done
Sequenced



1



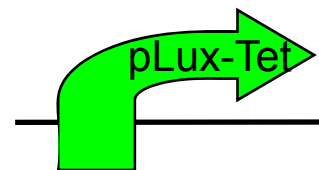
K176000 X+P with nicks
Sequence OK



P1010 in pSB1A3 in DB 3.1
Length OK

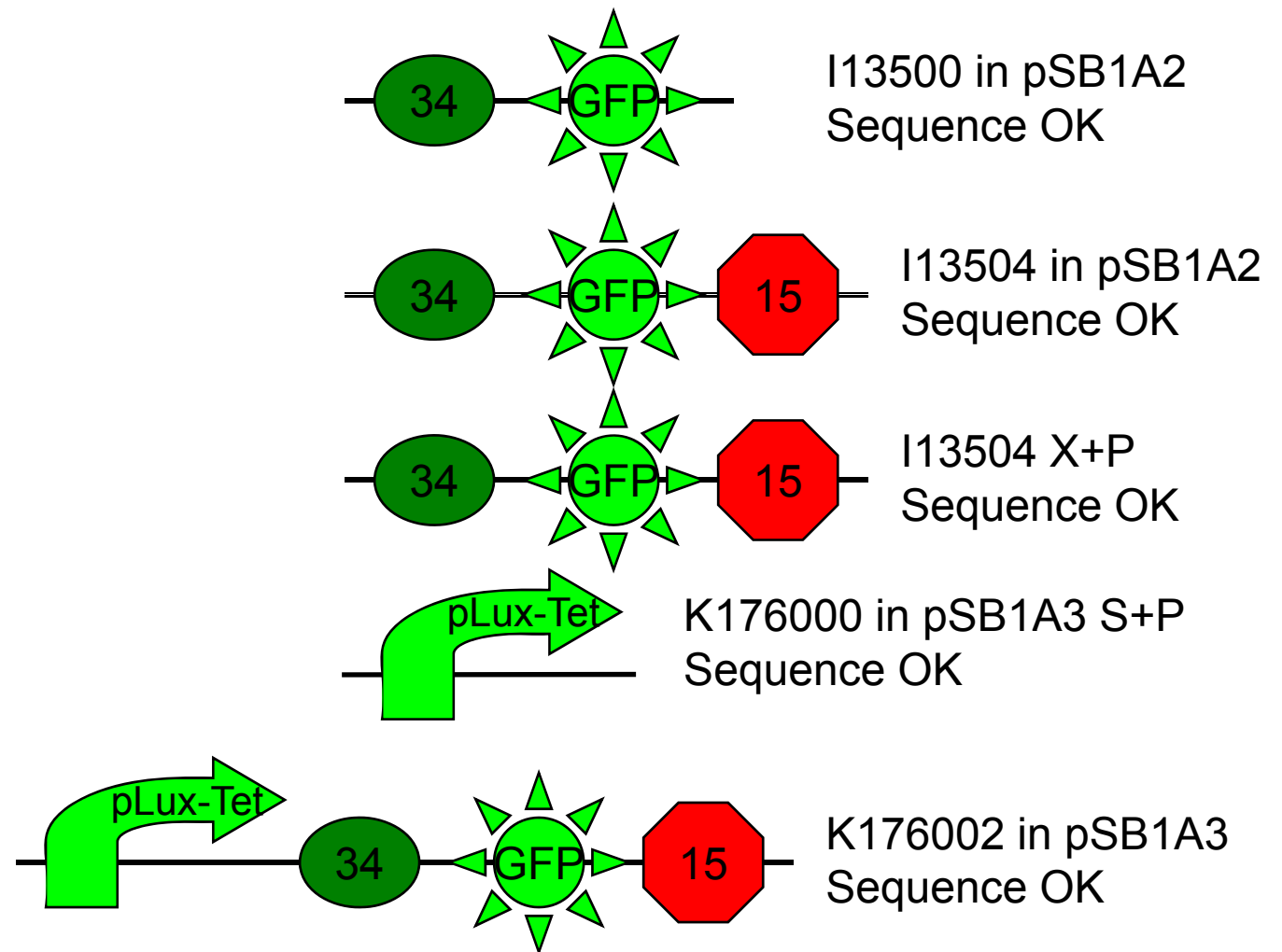


pSB1A3 X+P
Length OK



K176000 in pSB1A3
Sequence OK

2

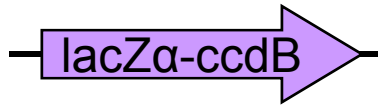


4



K145151 in pSB1A2 in DB 3.1

Sequence OK



K176003 Fragment Length OK



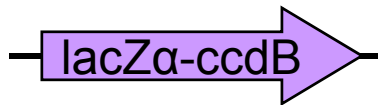
B0034 in pSB1A2
Sequence OK



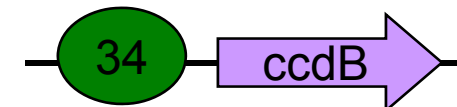
B0034 in pSB1A2 S+P
Length OK



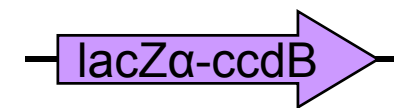
K145151 X+P
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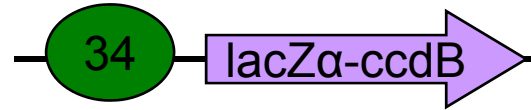
K176003 X+P
Length OK



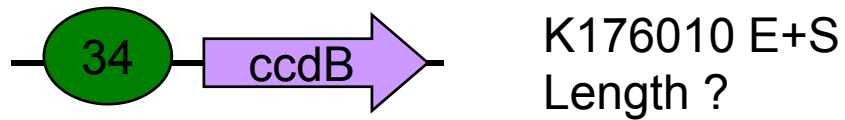
K176010 in pSB1A2 in DB 3.1
Sequencing



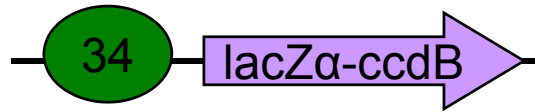
K176003 in pSB1A3 in DB 3.1
Sequence OK



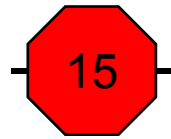
K176027 in pSB1A2 in DB 3.1
Sequencing



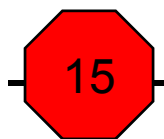
K176010 E+S
Length ?



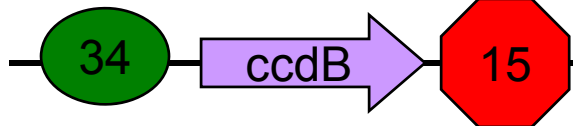
K176027 E+S
Length OK



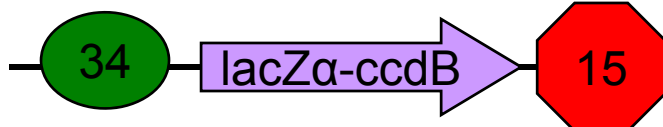
B0015 in pSB1AK3
Sequence OK



B0015 in pSB1AK3 E+X
Length OK

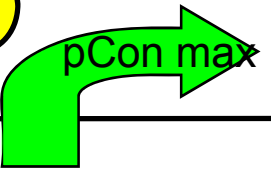


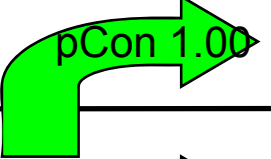
in pSB1AK3 in DB 3.1
Ligation

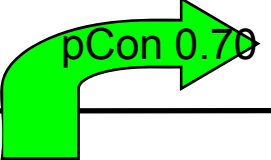


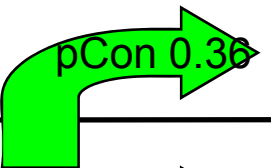
in pSB1AK3 in DB 3.1
Ligation

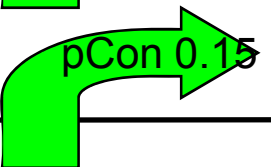
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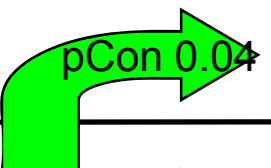
 pCon max J23119 in pSB1A2
Sequence OK

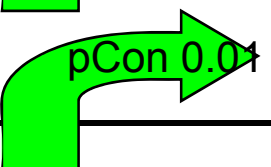
 pCon 1.00 J23100 in J61002
Sequence ?

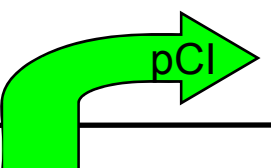
 pCon 0.70 J23101 in J61002
Sequence OK

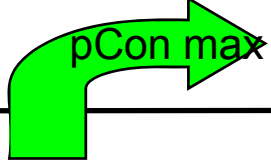
 pCon 0.36 K176009 in J61002
Sequence OK

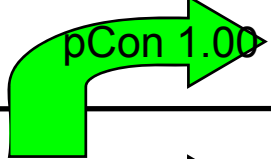
 pCon 0.15 K176008 in J61002
Sequence OK

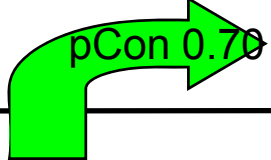
 pCon 0.04 J23109 in J61002
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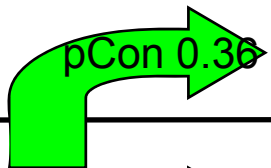
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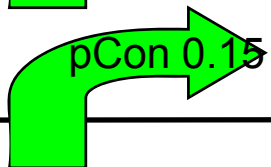
 pCl R0051 in pSB1A2
Sequence ?

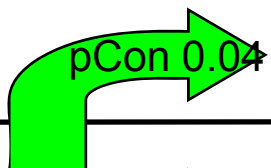
 pCon max J23119 X+P
Sequence OK

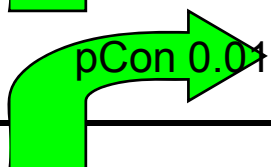
 pCon 1.00 J23100-J61002SF X+P
Sequence ?

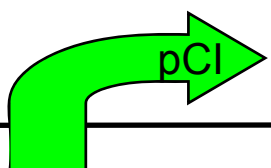
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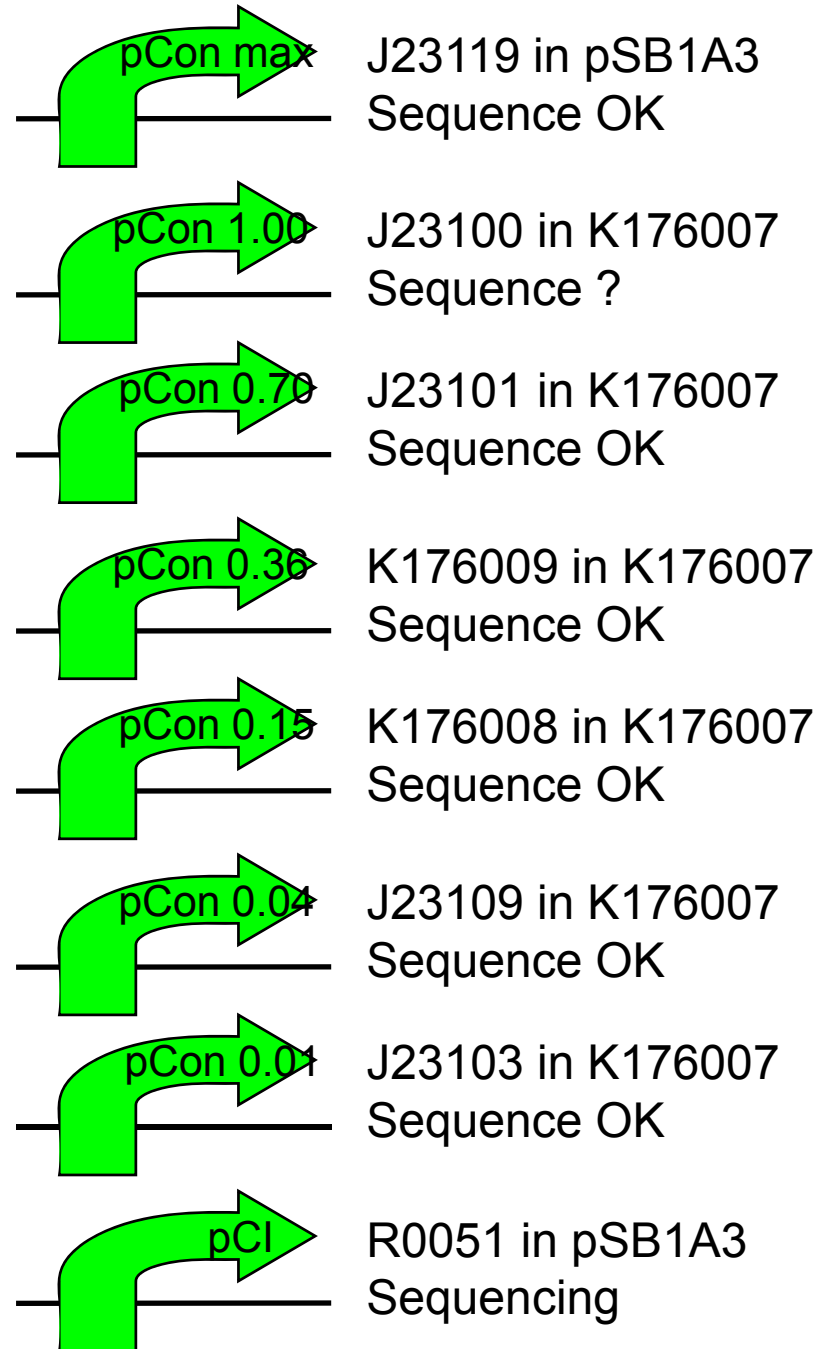
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Sequence OK

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Sequence OK

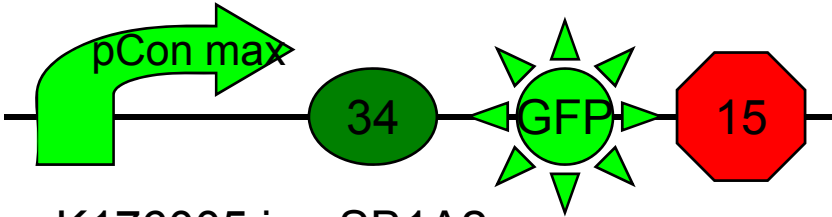
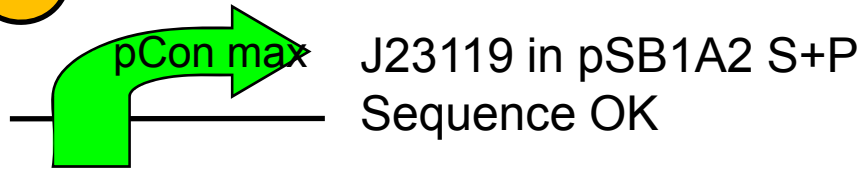
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Sequence OK

 pCon 0.04 J23103-J61002SF X+P
Sequence OK

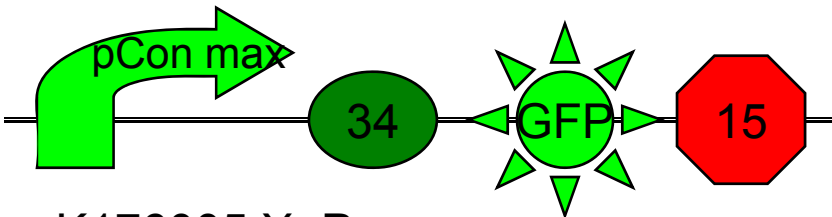
 pCl R0051 X+P
Length ?



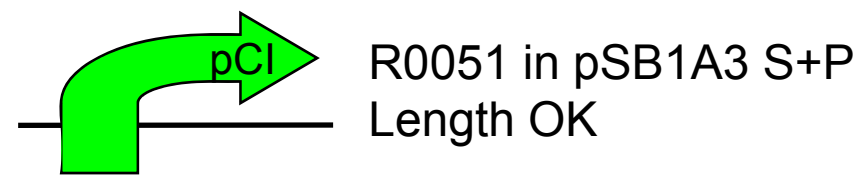
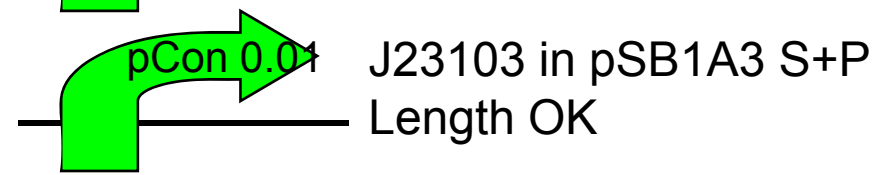
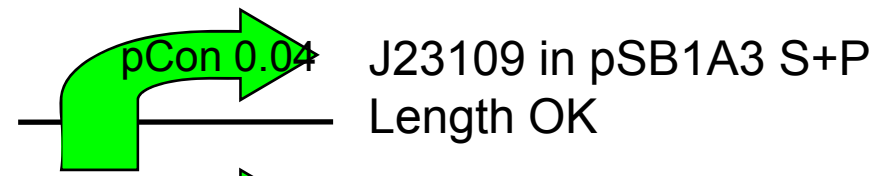
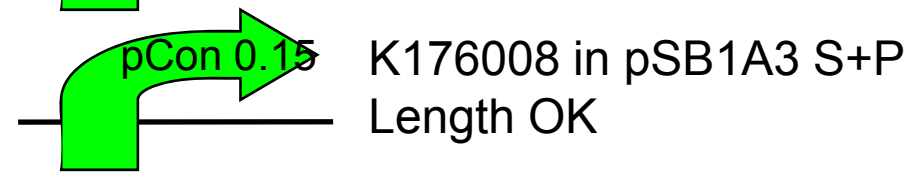
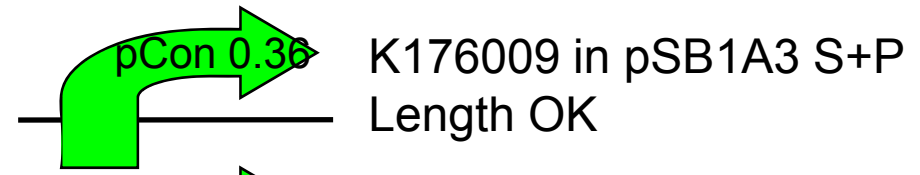
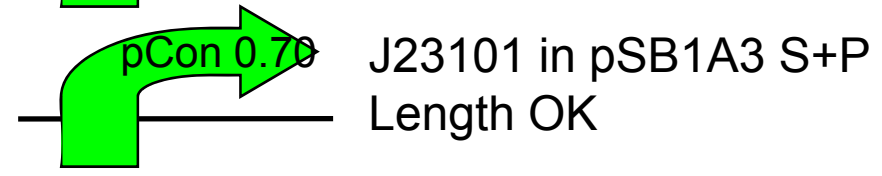
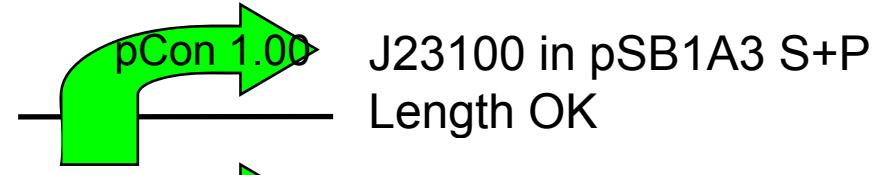
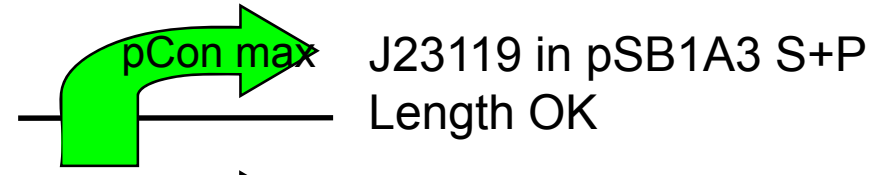
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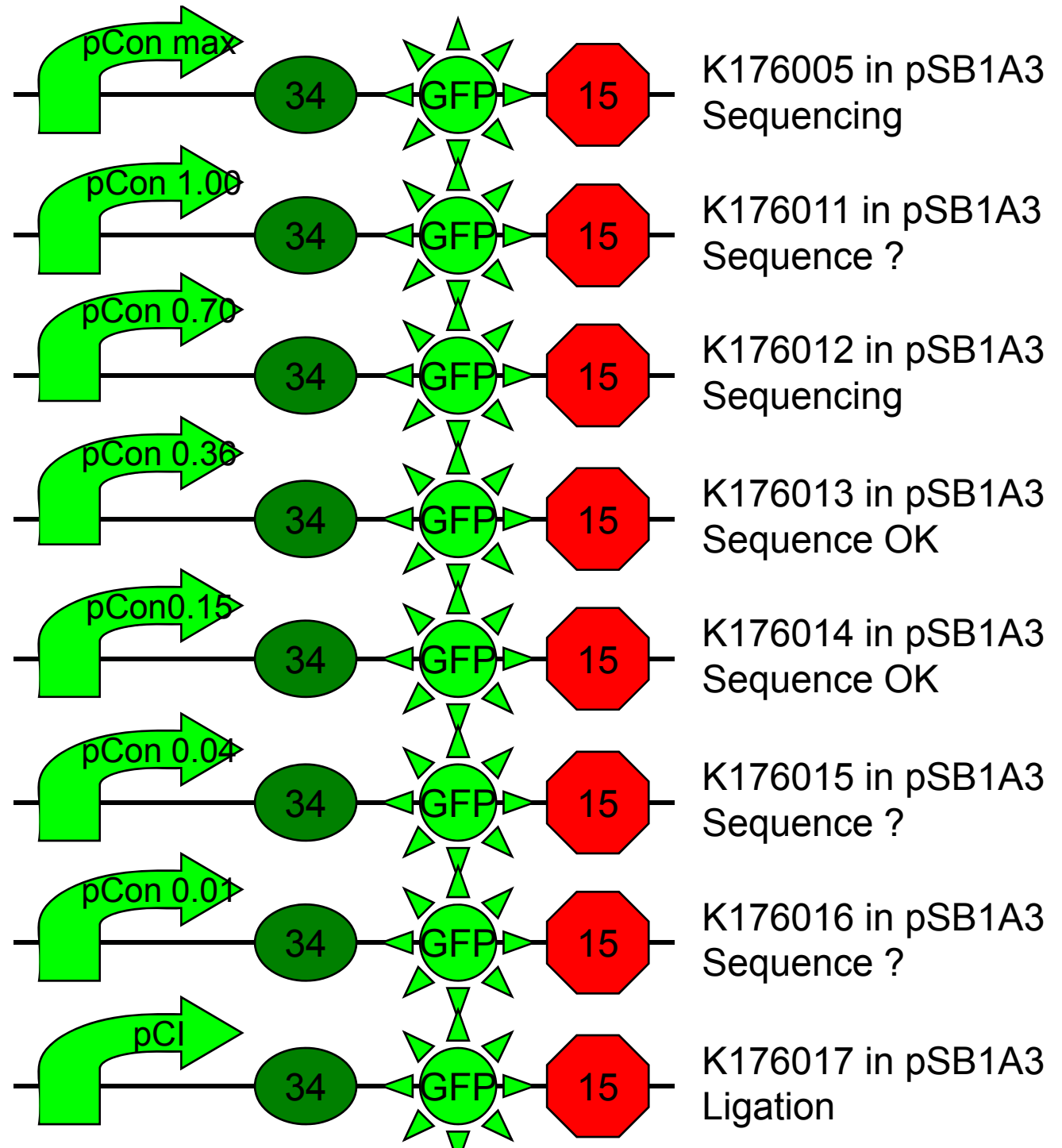


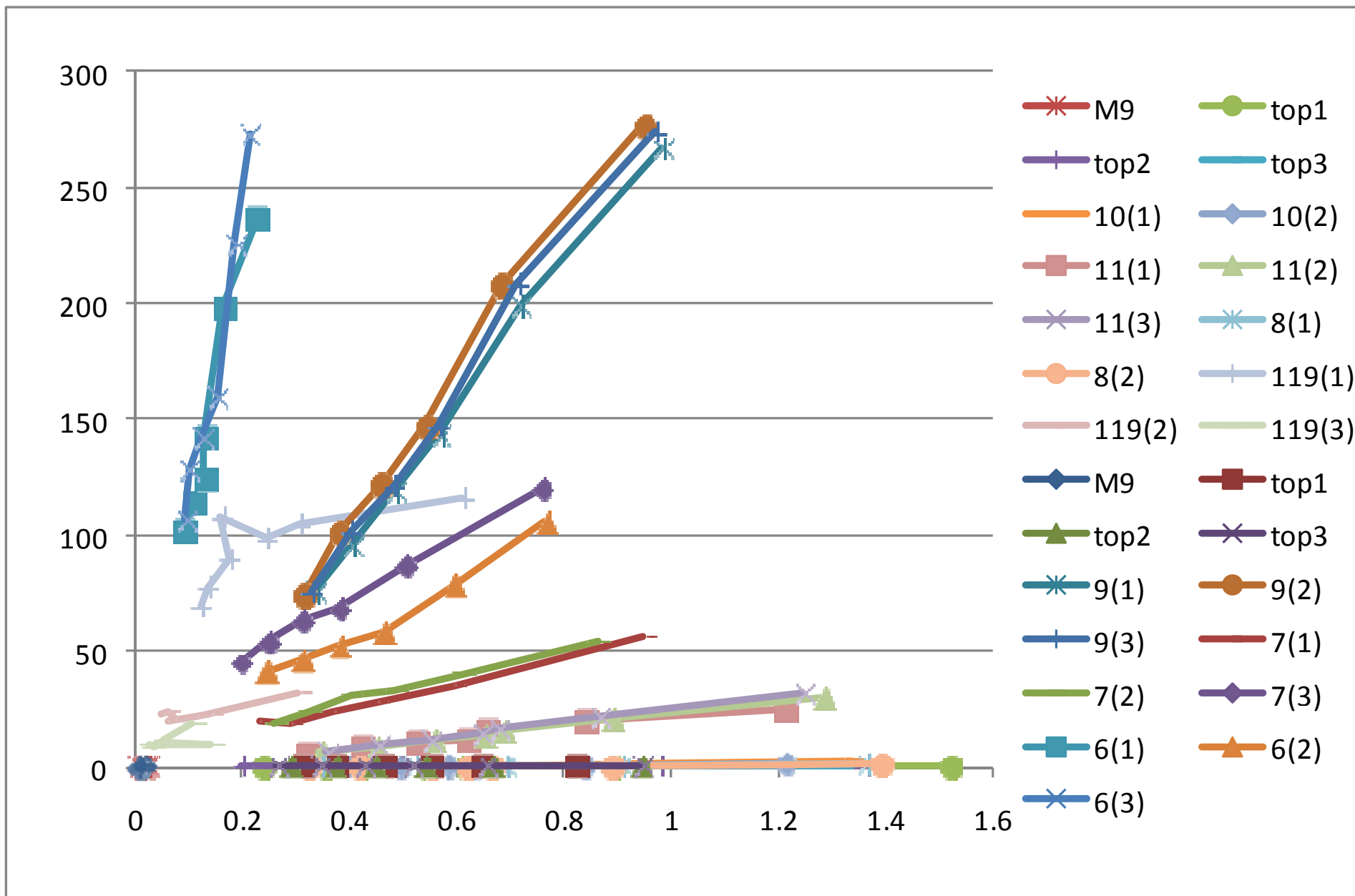
K176005 in pSB1A2
Sequence OK



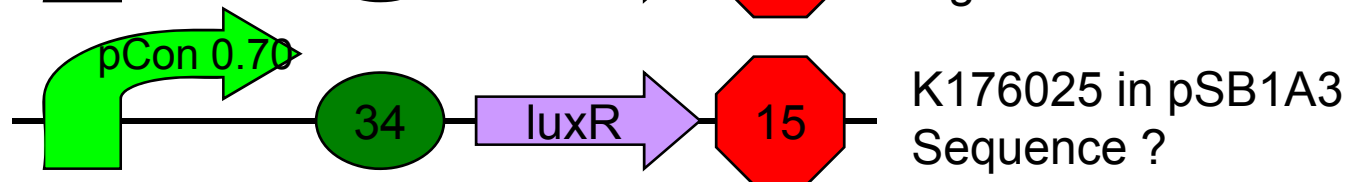
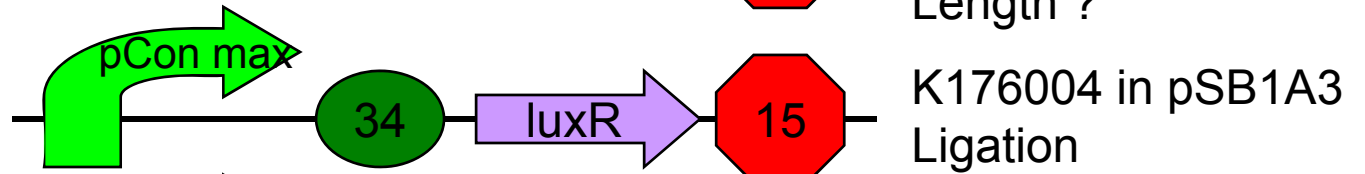
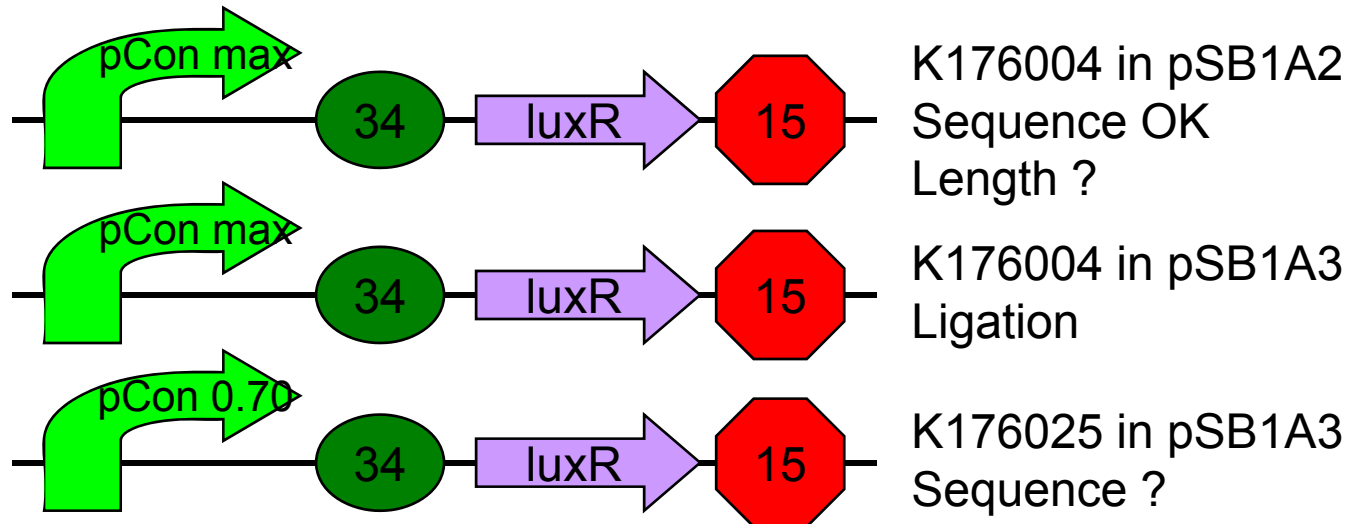
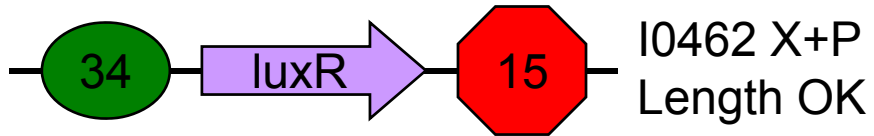
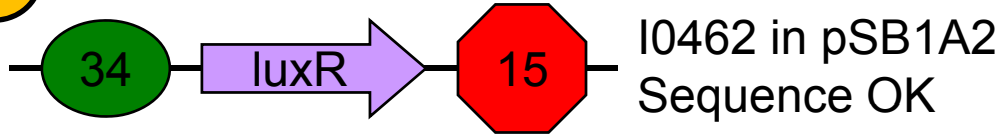
K176005 X+P
Length OK







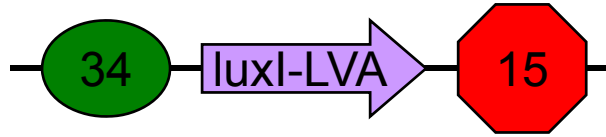
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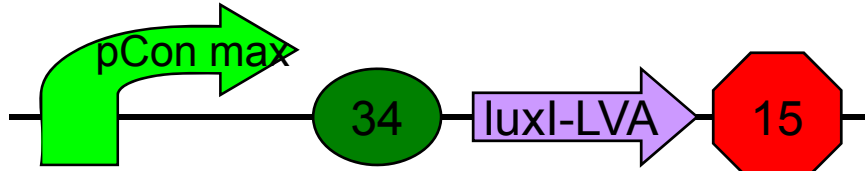
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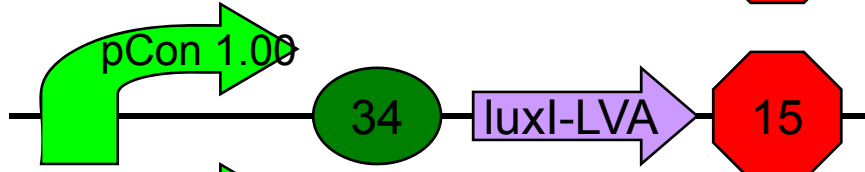
K082014 X+P
Length OK



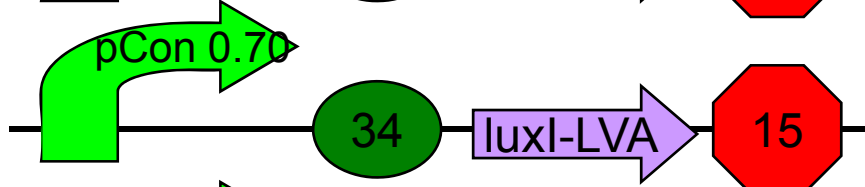
K082014 in pSB1AK3
Sequence OK



K176018 in
pSB1A3
Mini-Prep

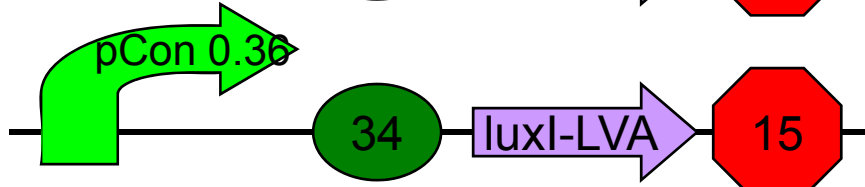


K176019 in
pSB1A3
Sequence OK

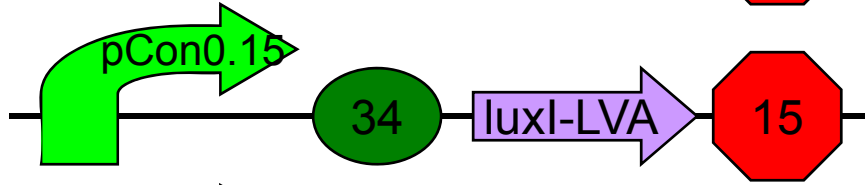


K176020 in
pSB1A3
Sequence ?

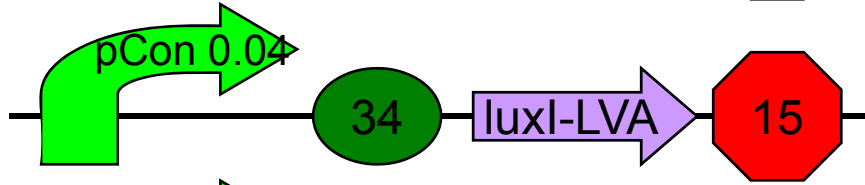
Measurement



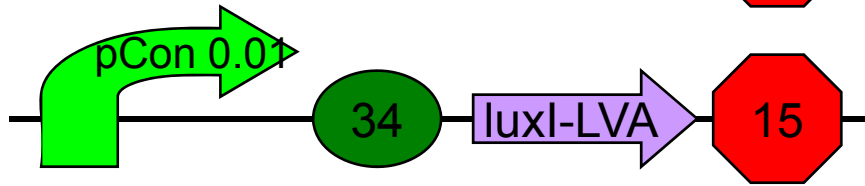
K176021 in
pSB1A3
Sequence OK



K176022 in
pSB1A3
Sequence OK

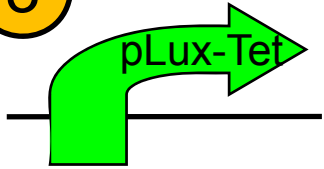


K176023 in
pSB1A3
Sequence OK

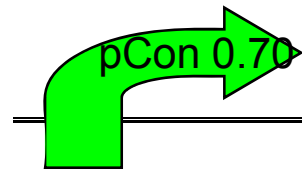


K176024 in
pSB1A3
Sequence OK

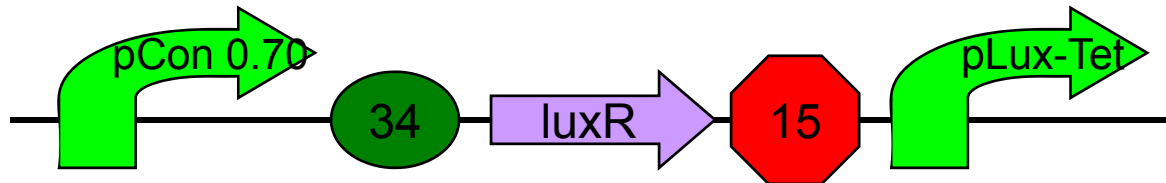
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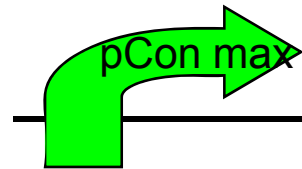
K176000 in pSB1A3 E+X
Length OK



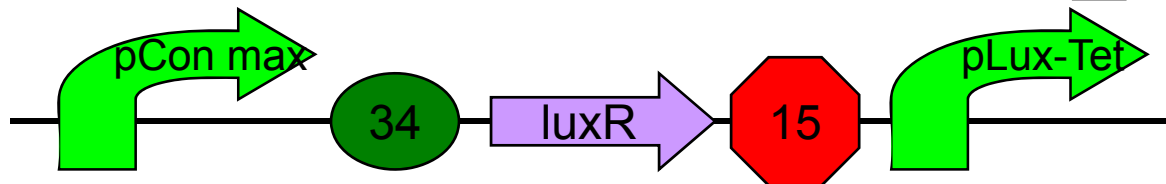
K176025 E+S
Length OK



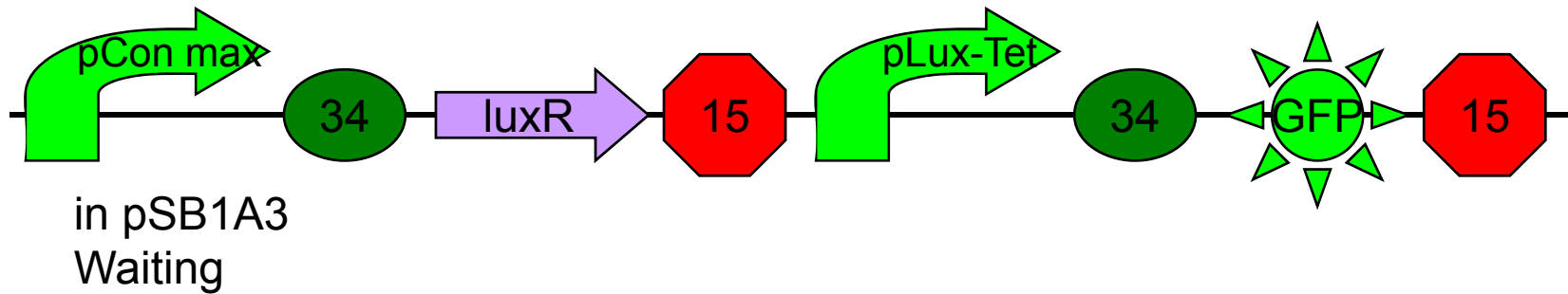
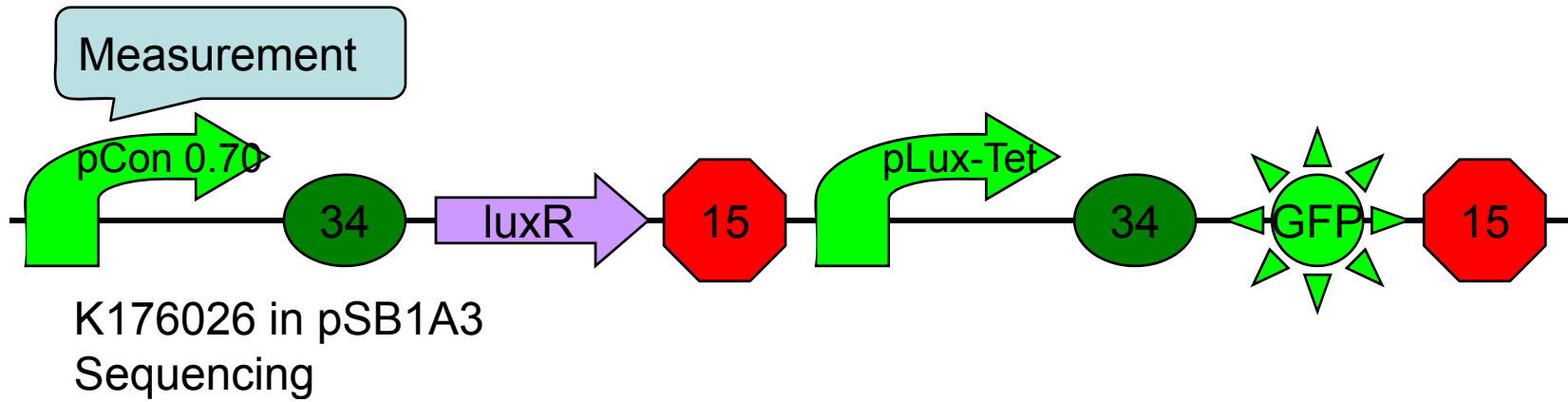
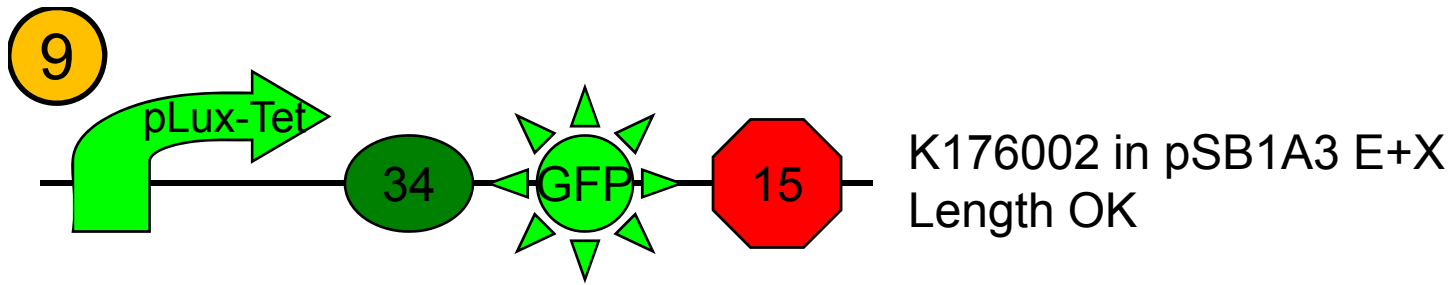
K176028 in pSB1A3
Ligation

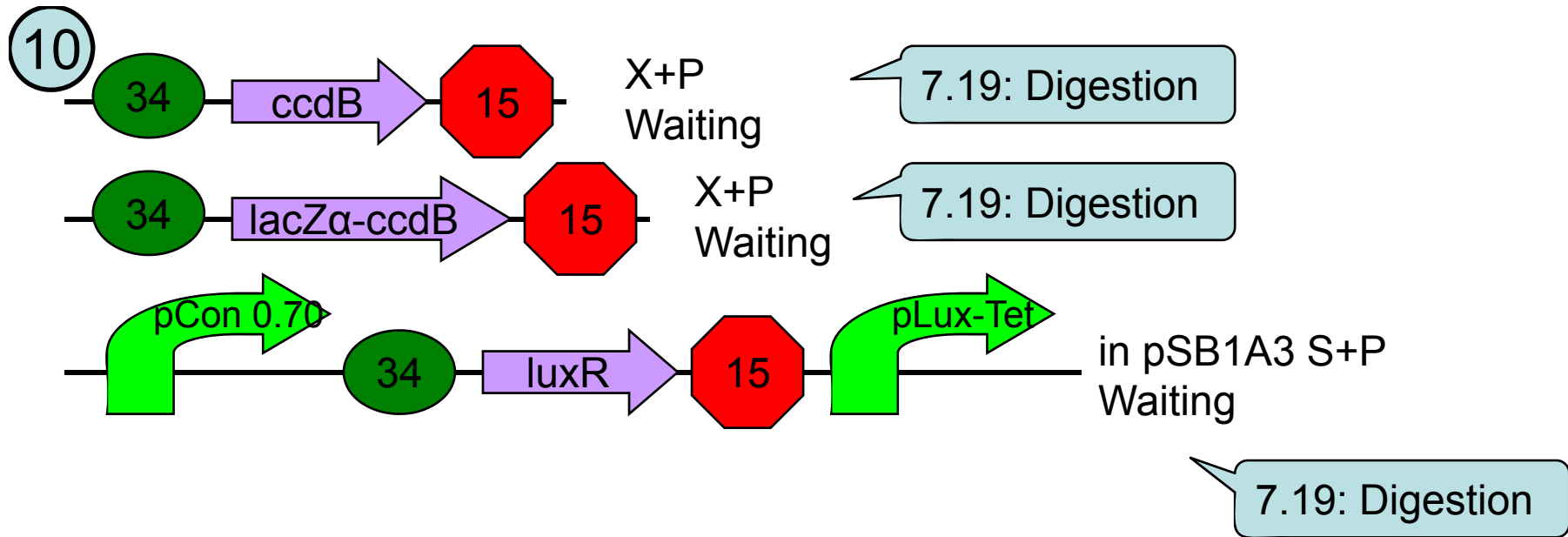


K176004 E+S
Waiting

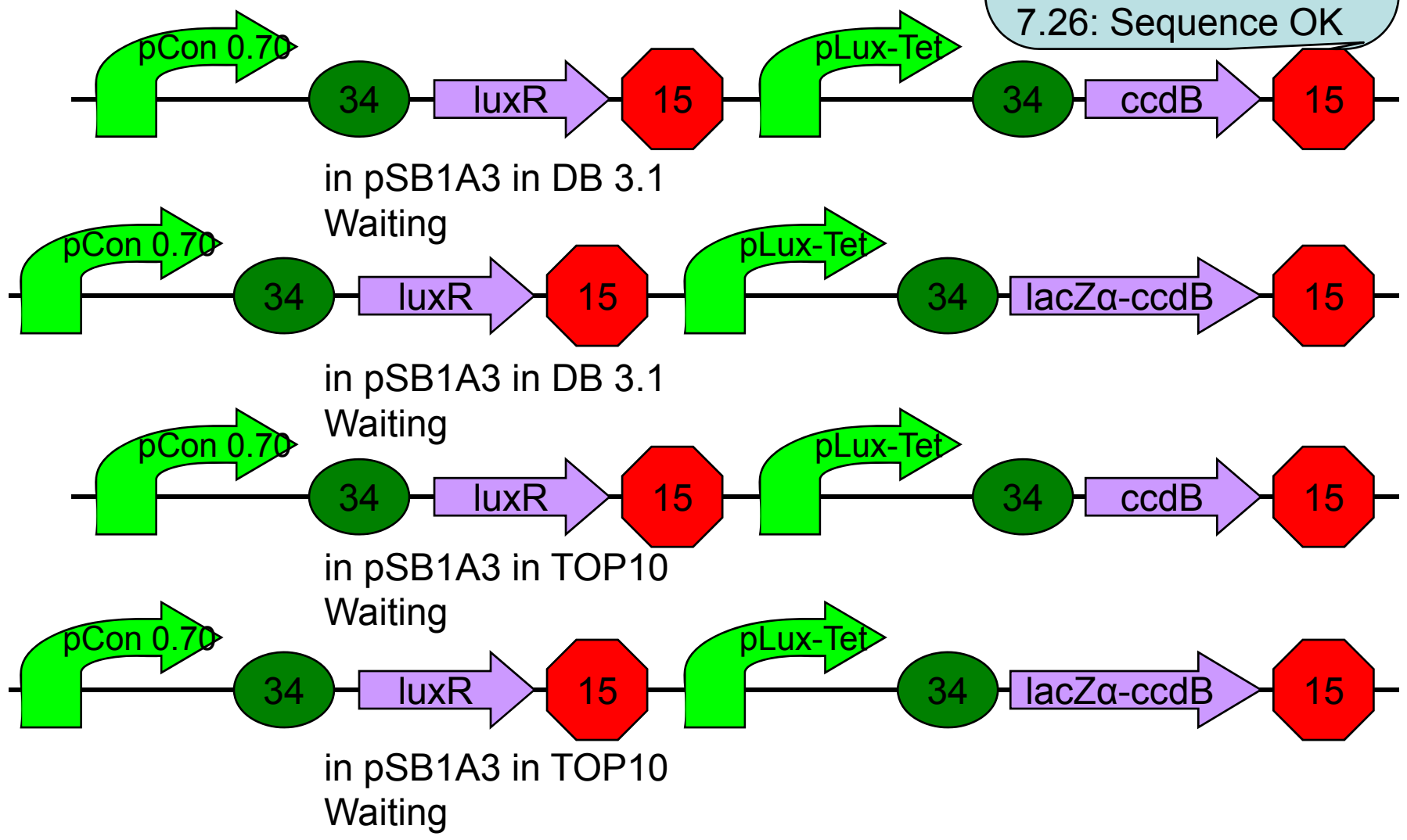


in pSB1A3
Waiting

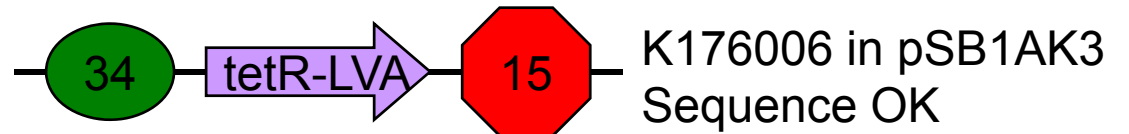
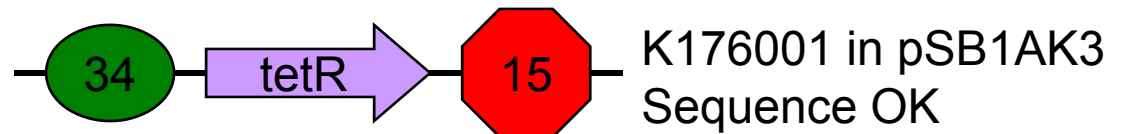
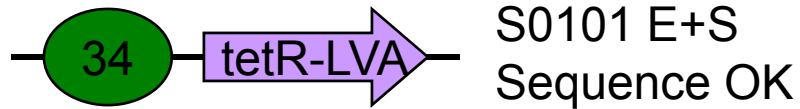
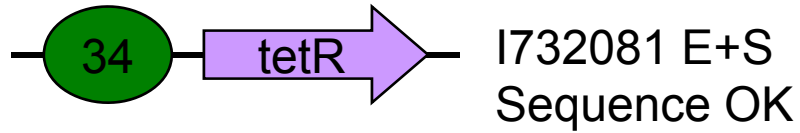
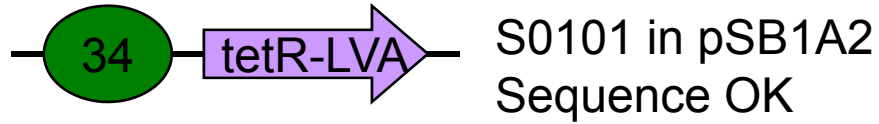
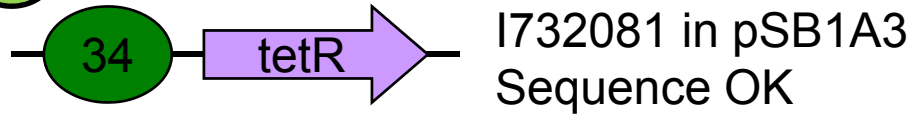




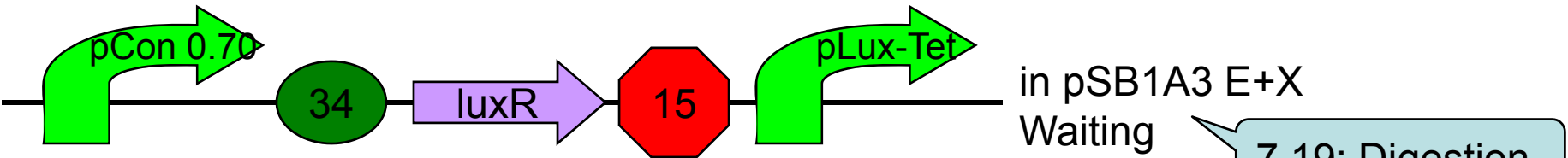
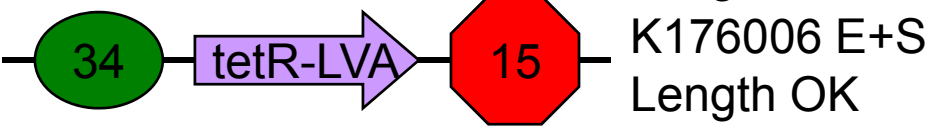
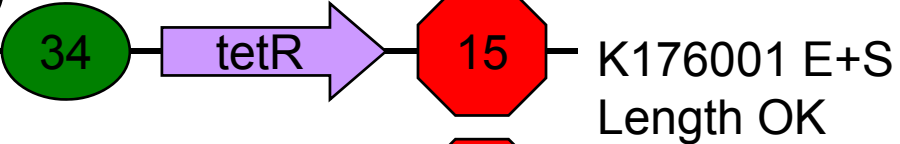
7.18: Competent Cells
 7.20: Ligation
 7.21: Colony PCR;
 Mini-Prep
 7.22: Sequencing
 7.23: Measurement
 7.26: Sequence OK



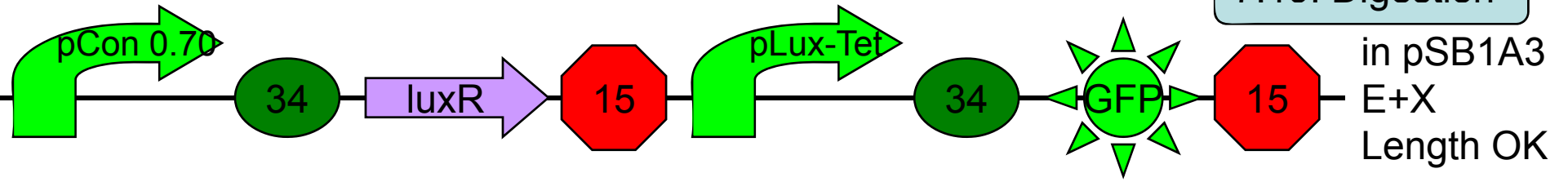
11

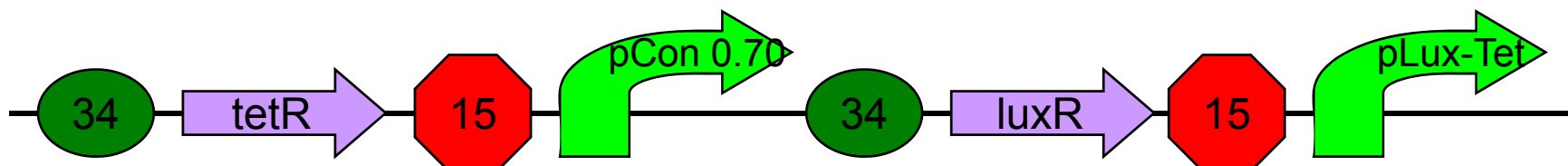


12

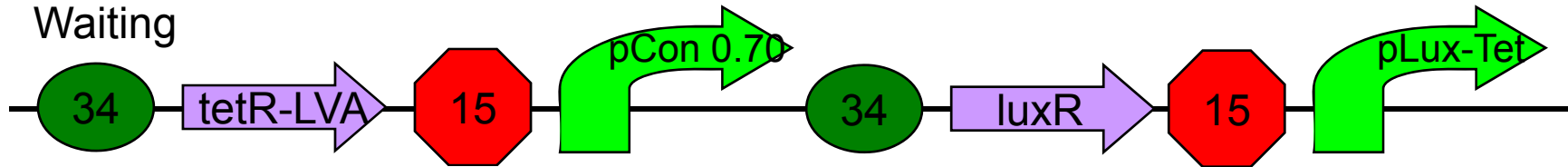


7.19: Digestion

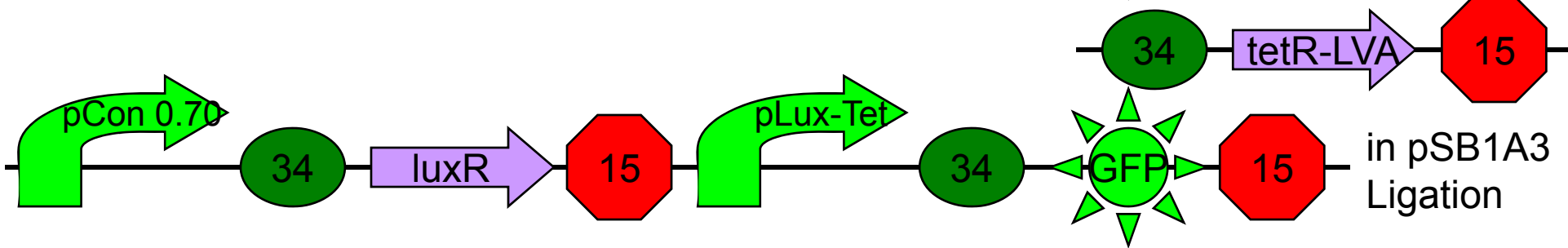
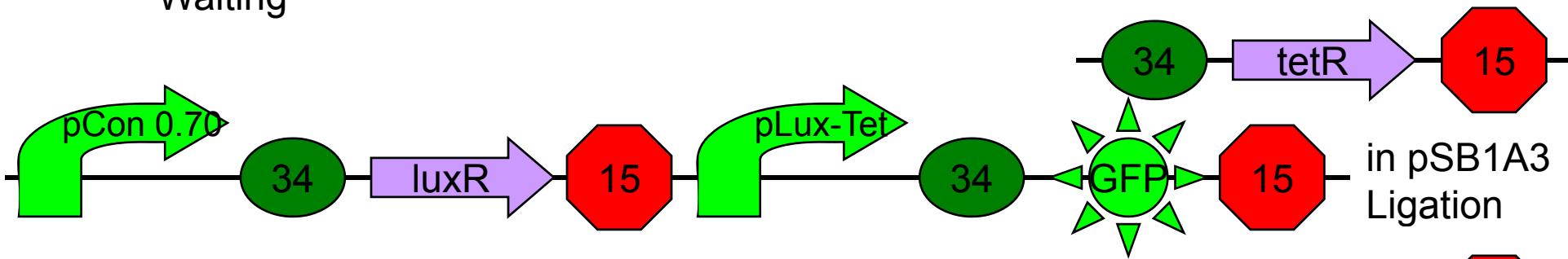




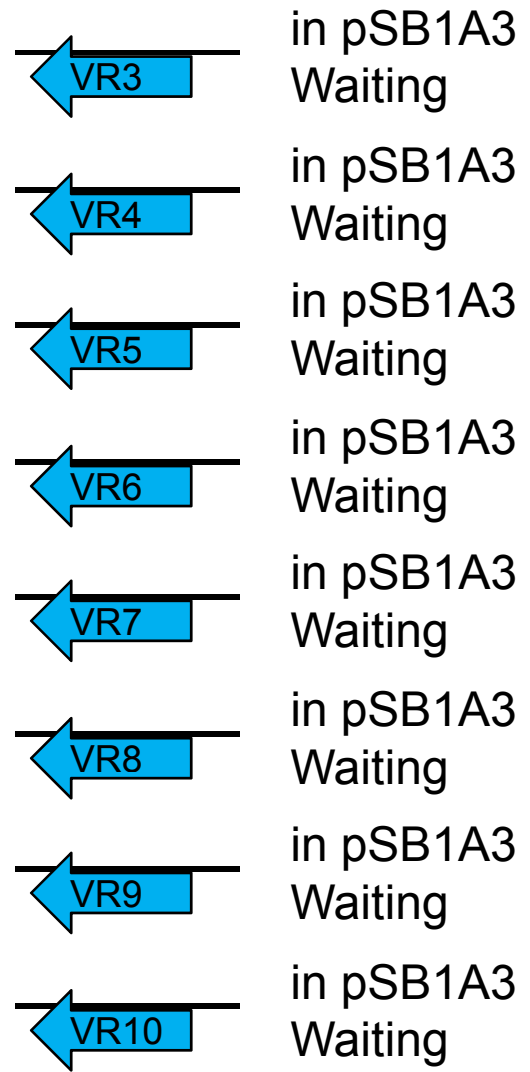
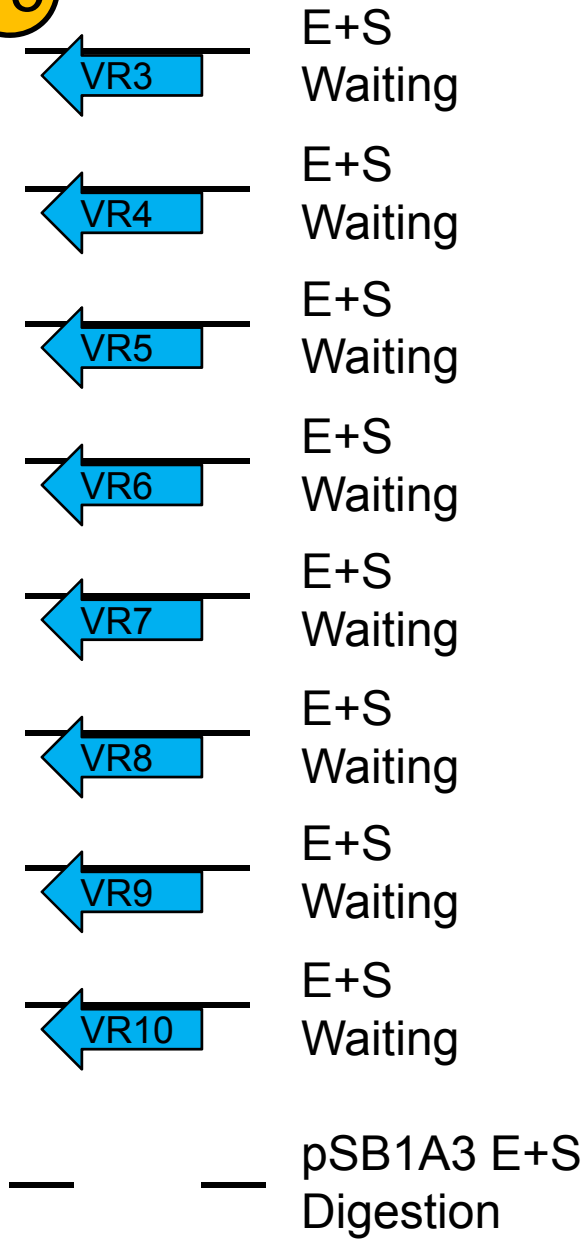
in pSB1A3
Waiting



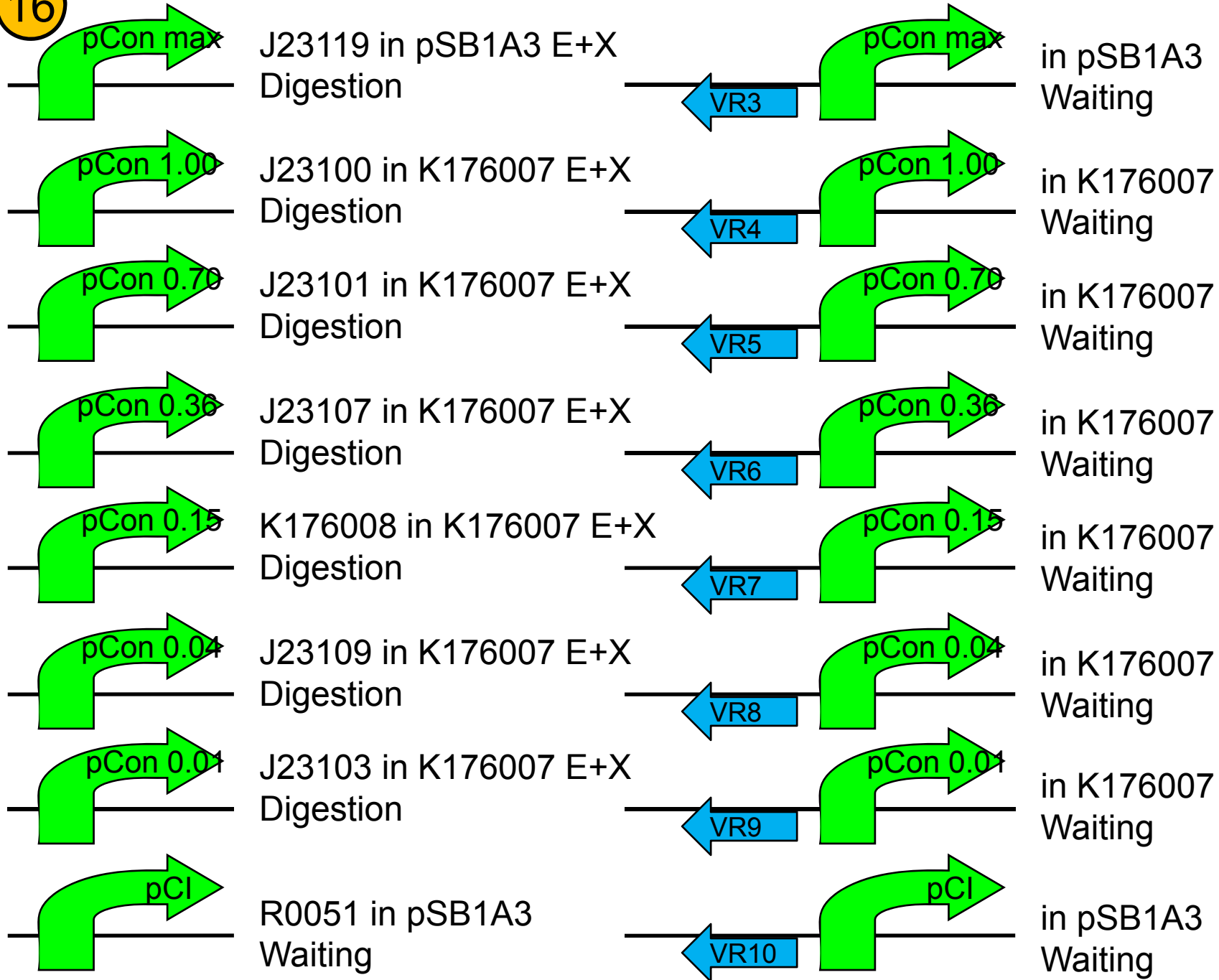
in pSB1A3
Waiting



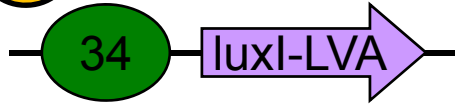
15



16



20



C0261 in pSB1A2
Sequence OK

Favorite USTC 2009 iGEM Team Parts

[Edit](#)

-?-	Name	Type	Description	Designer	Length
-----	------	------	-------------	----------	--------

USTC 2009 iGEM Team Parts Sandbox

[Edit](#)

-?-	Name	Type	Description	Designer	Length
	BBa_K176000	Regulatory	pLux/Tet Hybrid Promoter: (LuxR+,TetR-)->PoPS	Danqian Liu, Chao Li, Hao Jiang	72
	BBa_K176001	Generator	PoPS->RBS+TetR(without LVA)+T	Chao Li,Danqian Liu,Hao Jiang	782
	BBa_K176002	Reporter	pLux/Tet(K176000)(LuxR+,TetR-)->RBS+GFP+T	Chao Li,Danqian Liu,Hao Jiang	955
	BBa_K176003	Coding	lacZalpha-ccdB coding sequence	Zongxiao He, Hao Jiang	480
	BBa_K176004	Generator	pCon max(J23119)->RBS+LuxR+T	Chao Li,Danqian Liu,Hao Jiang	979
	BBa_K176005	Reporter	pCon max(J23119)->RBS+GFP+T	Chao Li,Danqian Liu,Hao Jiang	918
	BBa_K176006	Generator	PoPS->RBS+TetR(with LVA)+T	Chao Li,Danqian Liu,Hao Jiang	840
W	BBa_K176007	Plasmid_Backbone	pSB1A3 with the suffix of J61002 (mRFP)	Hao Jiang, Danqian Liu, Chao Li	3026
	BBa_K176008	Regulatory	constitutive promoter family member J23115 actual sequence	Hao Jiang, Danqian Liu, Chao Li	35
	BBa_K176009	Regulatory	constitutive promoter family member J23107 actual sequence	Hao Jiang, Danqian Liu, Chao Li	35
	BBa_K176010	Translational_Unit	PoPS->RBS+ccdB->PoPS	Zongxiao He, Hao Jiang	324
	BBa_K176011	Reporter	pCon 1.00(J23100)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
	BBa_K176012	Reporter	pCon 0.70(J23101)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
	BBa_K176013	Reporter	pCon 0.36(K176009)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
	BBa_K176014	Reporter	pCon 0.15(K176008)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
	BBa_K176015	Reporter	pCon 0.04(J23109)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
	BBa_K176016	Reporter	pCon 0.01(J23103)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
	BBa_K176017	Reporter	pCI(R0051)(lambda CI)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	932
	BBa_K176018	Signalling	pCon max(J23119)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
	BBa_K176019	Signalling	pCon 1.00(J23100)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
	BBa_K176020	Signalling	pCon 0.70(J23101)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
	BBa_K176021	Signalling	pCon 0.36(K176009)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
	BBa_K176022	Signalling	pCon 0.15(K176008)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
	BBa_K176023	Signalling	pCon 0.04(J23109)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
	BBa_K176024	Signalling	pCon 0.01(J23103)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841

Measurement

- General Conditions
 - Medium
 - LB
 - M9
 - Minimal
 - Supplemented
 - EZ Rich Define
 - pH-buffered TBK
 - pH-buffered LBK
 - Temperature
 - 37°C
 - 30°C
 - 34°C
 - Pre-warm
 - Shake
 - Dilution
 - Wash

http://openwetware.org/wiki/M9_medium

http://openwetware.org/wiki/M9_medium/minimal

http://openwetware.org/wiki/M9_medium/supplemented

http://openwetware.org/wiki/Neidhardt_EZ_Rich_Defined

<http://www.genome.wisc.edu/resources/protocols/ezmedium.htm>

GFP (PoPS)

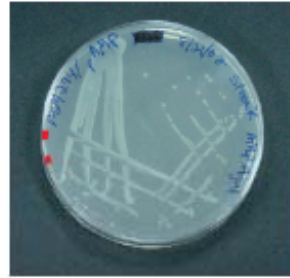
- Fluorospectrophotometer
- Plate Reader
- Flow Cytometer (FACS)
- Blotting
 - Northern
 - Western
- Realtime RT-PCR

- <http://partsregistry.org/Measurement>
- http://openwetware.org/wiki/The_BioBricks_Foundation:Standards/Technical/Measurement
- http://openwetware.org/wiki/Standardized_GFP_quantification
- Engineering the interface between cellular chassis (Barry Canton PhD thesis)
- Applying engineering principles to the design and construction of transcriptional devices (Reshma P. Shetty PhD thesis)

STEP 1: Streak 3 plates



A: TOP10
 B: BBa_I20260
 C: Your promoter!



STEP 2: Pick 3 colonies from each plate to start overnight cultures in Supplemented M9 Media at 37 C (9 tubes)



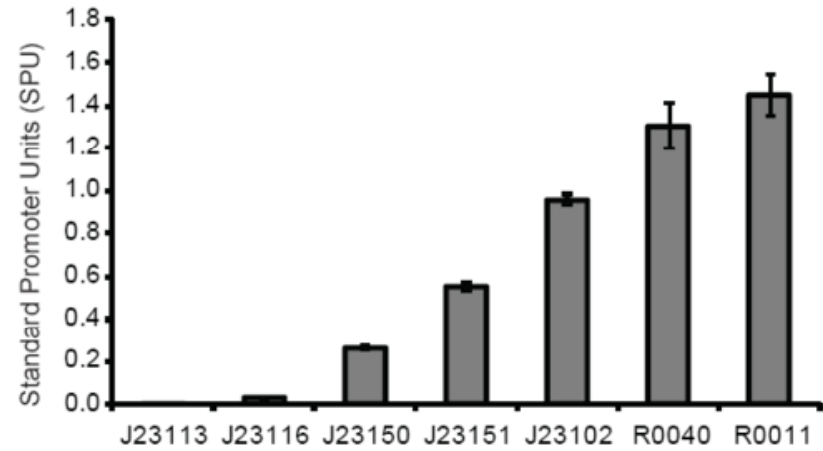
TOP10
 BBa_I20260
 Your Promoter



37C

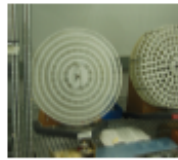
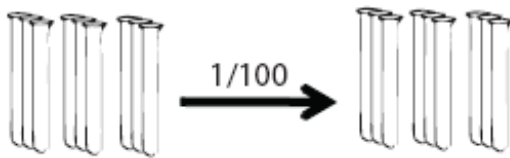


16 hours



Standard Promoter Units

STEP 3: Dilute 1/100 into fresh, pre-warmed media incubate at 37C (9 tubes)



37C

STEP 4: After 3 hours measure GFP and OD



3 hours



GFP

OD

STEP 5: After another half hour measure GFP and OD again



1/2 hour



GFP

OD

2006 Berkeley **J23100~J23119**
 Reported activities of the promoters are given as the relative fluorescence of these plasmids in strain TG1 grown in LB media to **saturation**. See part J61002 for details on their use.

AHL

- Rapid Screening of Quorum-Sensing Signal N-Acyl Homoserine Lactones by an In Vitro Cell-Free Assay
- Detection of N-acylhomoserine lactones in lung tissues of mice infected with *Pseudomonas aeruginosa*
- Detecting and characterizing N-acyl-homoserine lactone signal molecules by thin-layer chromatography
- Detection of N-acyl homoserine lactones using a *traI-luxCDABE*-based biosensor as a high-throughput screening tool
- On-line high-performance liquid chromatography-mass spectrometric detection and quantification of N-acylhomoserine lactones, quorum sensing signal molecules, in the presence of biological matrices
- Detection of quorum-sensing N-acyl homoserine lactone signal molecules by bacterial biosensors

CcdB & LacZ α

- Programmed population control by cell–cell communication and regulated killing
- A synthetic *Escherichia coli* predator–prey ecosystem
- LacZ α
 - X-gal
 - ONPG
 - <http://parts.mit.edu/igem07/index.php/USTC/BetaGalactosidaseAssay>

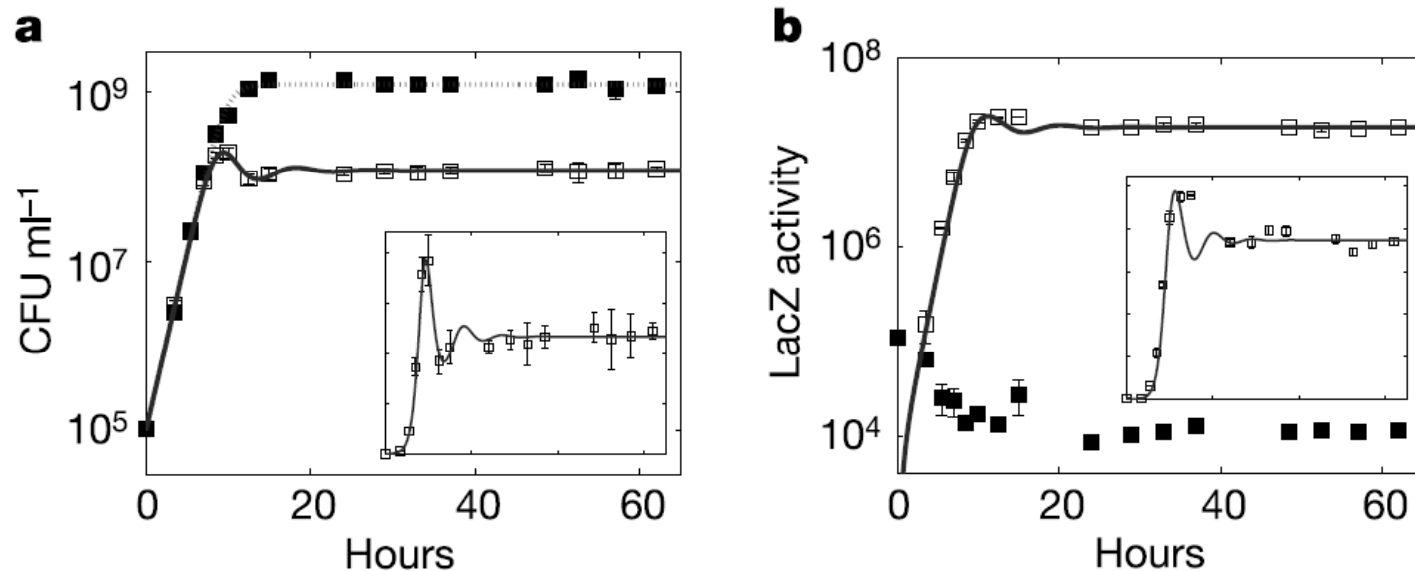


Figure S2:

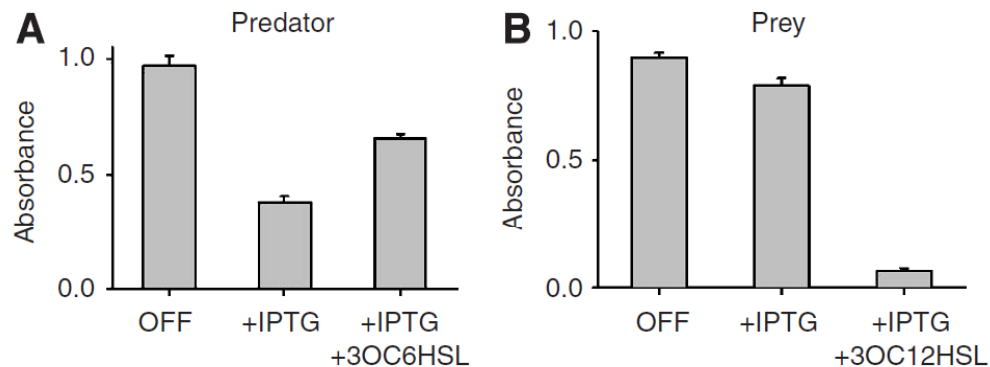
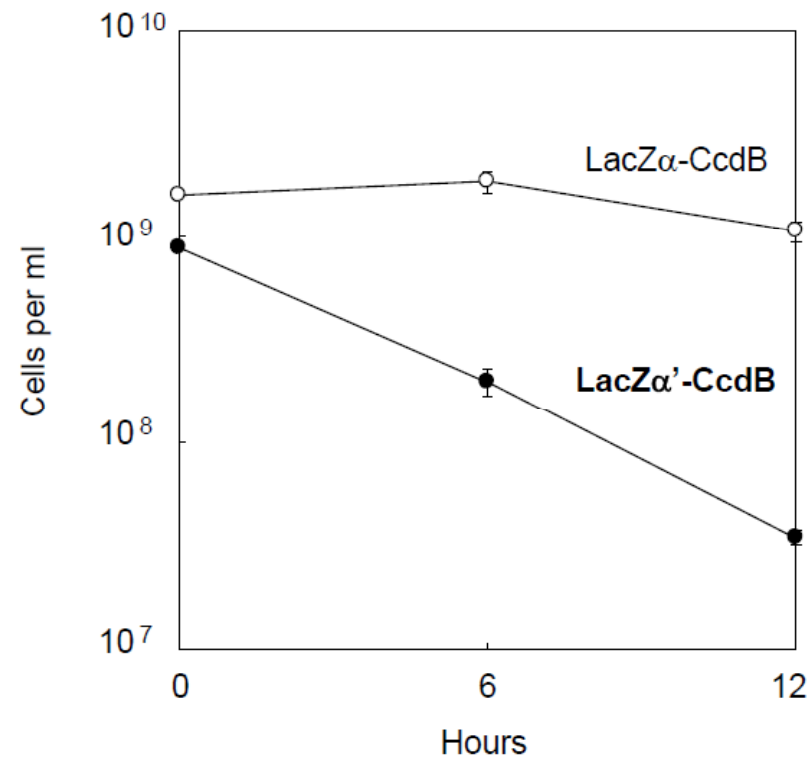


Figure 1 Individual growth behaviors (without interactions) of **(A)** predator and **(B)** prey cells in liquid media. For each condition, 6 ml LBK medium containing chloramphenicol and kanamycin was inoculated with a single bacterial colony and was divided into three 2 ml cultures: 'OFF' cultures contained no inducers, '+ IPTG' cultures contained 1 mM IPTG and '+ IPTG + AHL' contained 1 mM IPTG and 100 nM AHL, respectively. After 20h of incubation (light gray bars), optical densities (ODs) of these cultures were measured with a microplate reader (see Supplementary information). Error bars represent standard deviation of triplicate cultures.



Wiki

- Team project description
- Notebook
 - Meetings
 - Lab Work
 - Sample Naming Sheets
- Other
 - Team
 - Project
 - Parts
 - Modeling
 - Human Practice



2009

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Calendar of Events

IGEM 2009 Calendar of Events

Note: Dates in grey have not been finalized yet. Make sure to check the calendar periodically for any changes!

- | | |
|----------------|---|
| 19 February | IGEM 2009 registration opens |
| 31 March | IGEM 2008 registration closes; Team registration fee due |
| 13 May | DNA Distribution sent to teams (target deadline; subject to change) |
| 16/17 May | iGEM Workshop, MIT, USA |
| 1 June | Visa invitation letter requests due |
| 20/21 June | iGEM Workshop, Europe |
| 27/28 June | iGEM Workshop, Asia |
| 15 June | Preliminary team rosters due |
| 1 August | Team project descriptions due |
| TBD | Jamboree attendance fee due |
| TBD | Request for variance due (notice and description of any use of non-standard parts or devices schemes due) |
| | Track selection due |
| | Project abstracts due |
| TBD | Team rosters due |
| TBD | Project and part documentation due, including documentation for all medal criteria |
| | BioBrick Part DNA needs to be received by the Registry |
| 30 Oct - 2 Nov | IGEM Competition Jamboree, MIT, USA |



USTC

University of Science and Technology of China

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Team:USTC/Notebook

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[1 Team:USTC/Notebook](#)[1.1 Meetings](#)[1.2 Brainstorming](#)[1.3 Lab Work](#)[1.4 Sample Naming Sheets](#)

Meetings

All the reports information, slides and audio records of our meetings are arranged here.

December	January	February	March	April	May
M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31
June	July	August	September	October	
M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30	M T W T F S S 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31	

			30 31		
June	July	August	September	October	
M T W T F S S	M T W T F S S	M T W T F S S	M T W T F S S	M T W T F S S	
1 2 3 4 5 6 7	1 2 3 4 5	1 2	1 2 3 4 5 6	1 2 3 4	
8 9 10 11 12 13 14	6 7 8 9 10 11 12	3 4 5 6 7 8 9	7 8 9 10 11 12 13	5 6 7 8 9 10 11	
15 16 17 18 19 20 21	13 14 15 16 17 18 19	10 11 12 13 14 15 16	14 15 16 17 18 19 20	12 13 14 15 16 17 18	
22 23 24 25 26 27 28	20 21 22 23 24 25 26	17 18 19 20 21 22 23	21 22 23 24 25 26 27	19 20 21 22 23 24 25	
29 30	27 28 29 30 31	24 25 26 27 28 29 30	28 29 30	26 27 28 29 30 31	
		31			

Brainstorming

- [2009-02-10](#)
- [2009-03-07](#)
- [2009-03-22](#)
- [2009-03-30](#)

Lab Work

We chose to use [iPad](#) as our Electronic Lab Notebook. iPad is recommended on OWW [\[1\]](#).

Our work progress is also updated [here](#).

May	June	July	August	September	October
M T W T F S S	M T W T F S S	M T W T F S S	M T W T F S S	M T W T F S S	M T W T F S S
1 2 3	1 2 3 4 5 6 7	1 2 3 4 5	1 2	1 2 3 4 5 6	1 2 3 4
4 5 6 7 8 9 10	8 9 10 11 12 13 14	6 7 8 9 10 11 12	3 4 5 6 7 8 9	7 8 9 10 11 12 13	5 6 7 8 9 10 11
11 12 13 14 15 16 17	15 16 17 18 19 20 21	13 14 15 16 17 18 19	10 11 12 13 14 15 16	14 15 16 17 18 19 20	12 13 14 15 16 17 18
18 19 20 21 22 23 24	22 23 24 25 26 27 28	20 21 22 23 24 25 26	17 18 19 20 21 22 23	21 22 23 24 25 26 27	19 20 21 22 23 24 25
25 26 27 28 29 30 31	29 30	27 28 29 30 31	24 25 26 27 28 29 30	28 29 30	26 27 28 29 30 31
			31		

Sample Naming Sheets

We chose to use [this wiki page](#) to manage samples. The names are generated with a program written by [Jian Zhan](#).



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Instructional Videos

Workshop videos: Videos from the iGEM 2009 spring workshop @ MIT are available below for streaming and download, and higher quality downloads will follow shortly. You can also visit our [iGEM channel @ blip.tv](#).

Welcome to iGEM



A short welcome and introduction to iGEM by Randy Rettberg

low high

Navigating the Registry



Reshma and Randy discuss navigating through [partsregistry.org](#)

low high

Introduction to Synthetic Biology



Tom Knight gives an introduction to parts based synthetic biology

low high

Project Ideas



Reshma Shetty gives suggestions on how teams may want to come up with project ideas

slides low high

Changes for iGEM 2009



Randy Rettberg discusses the changes that have taken place for iGEM 2009, the requirements for the teams, and judging the competition

low high

Standard Assembly



Reshma Shetty shows how parts on the registry are designed for standard assembly

slides low high

Promoters



Barry Canton discusses the promoter category of parts in the registry

low high

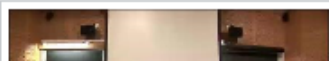
Making and Adding Parts



Reshma and Meagan show how to make and add parts to the registry

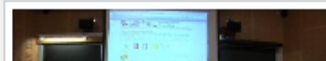
low high

Devices



Barry Canton discusses devices in the registry

Favorites and Shipping Parts



Meagan Lizarazo shows how to make your parts "Favorites" and ship them



using the registry



Measurements



Barry Canton discusses the importance of measuring and documenting the parts on the registry

2009 Distribution, QC, and Sequencing



Paul and Vinoo discuss an overview of the creation of the 2009 distribution and the quality control process. Randy discusses the sequencing tools on the registry.

Software Tools Track



Randy Rettberg discusses the software track for iGEM participants, as well as how software tools are integrated into the registry

Safety in iGEM



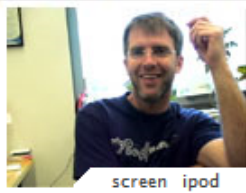
"What safety precautions should my team be taking while participating in iGEM? Why is this important?"

Drew Endy: Defining Synthetic Biology



"Make it easier to build things. Define the things you are building with by using standards. Hide biological complexity with abstraction."

Drew Endy: Believe in Synthetic Biology



"Why should you consider changing how you engineer biological systems from doing ad-hoc research to something that's a more scalable engineering framework?"













Drew Endy: What is a Standard Biological Part?



"What does it mean to have a Standard Biological Part - for example, a BioBrick-standard"








Episodes Archive

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[Syndicate Show](#) ▶

 <p>iGEM 2009 Spring Workshop: MIT Tom's talk on parts based synthetic biology</p>	 <p>iGEM 2009 Spring Workshop: MIT Randy's Welcome Speech</p>	 <p>Safety in iGEM</p>	 <p>Jam07 - Beginnings and Beyond</p>
 <p>Jam07 - From Challenge to Triumph</p>	 <p>Interview with Alja Oblak from the iGEM06 Ljubljana Team</p>	 <p>Brown iGEM07 Team - Introduction to iGEM</p>	 <p>Jam07 - What can synthetic biology do for you?</p>
 <p>Jam07 - Calgary - "Developing A Genetic Printer"</p>	 <p>Jam07 - Caltech - "Selection for Infection"</p>	 <p>iGEM 2007 Jamboree Good Times</p>	 <p>iGEM 2007 Jamboree Dance Off</p>

Episodes Archive

Back to show page ▶
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 <p>Jam07 - ETH Zurich - "educatETH E.coli System"</p>	 <p>tutorial 4.3 - Reviewing your part</p>	 <p>Tutorial 4.2 - Entering Part Sequence and Features</p>	 <p>Tutorial 4.1 - Adding and documenting a basic part</p>
 <p>iGEM Explainer 03 - Drew Endy: What is a Standard Biological Part</p>	 <p>iGEM Explainer 01 - Drew Endy: Defining Synthetic Biology</p>	 <p>iGEM Explainer 02 - Drew Endy: Believe in Synthetic Biology</p>	

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