E.ADEM v0.0.4.3

2009.7.11

2009.5.3

68 days

2009.7.11

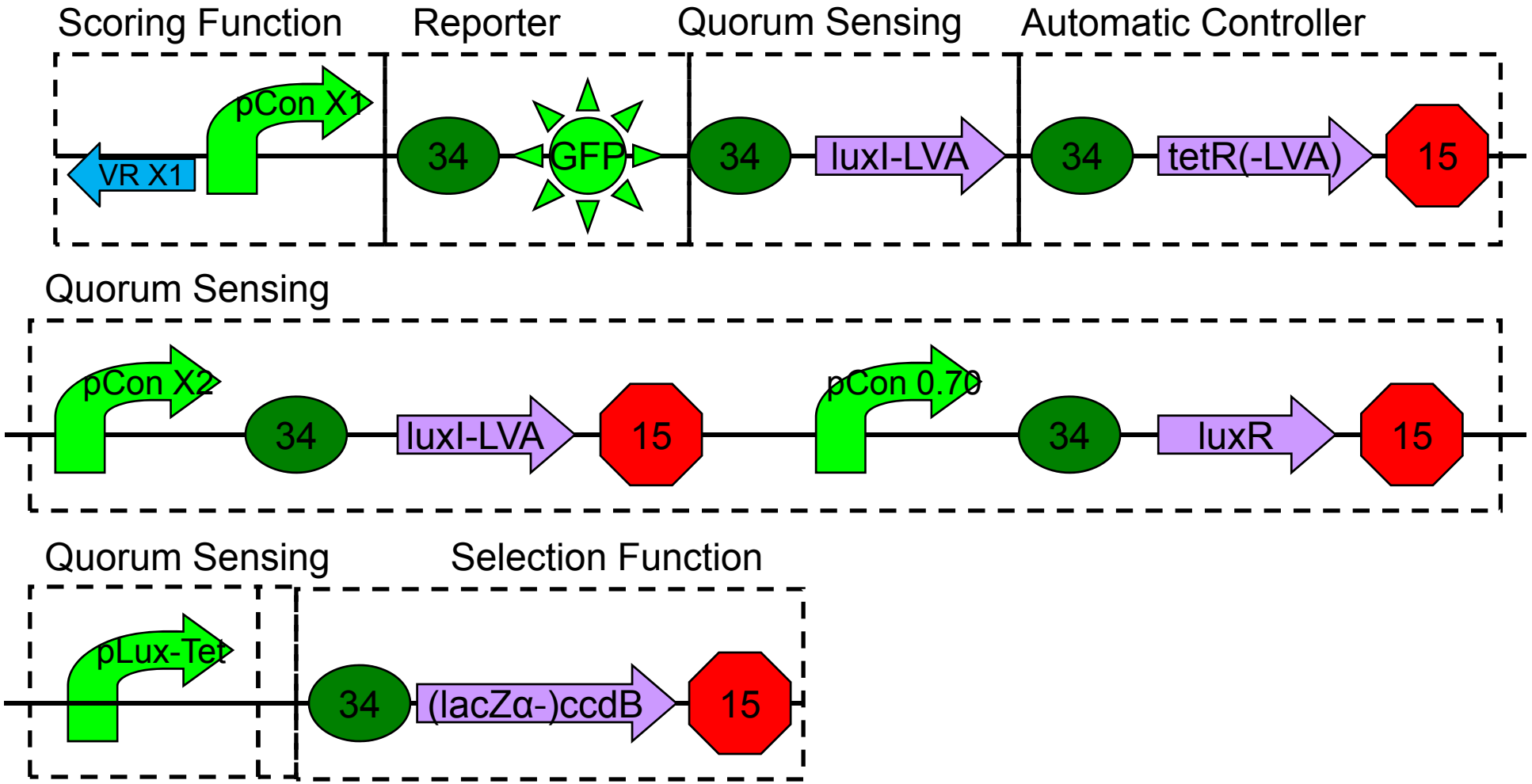
110 days

2009.10.30

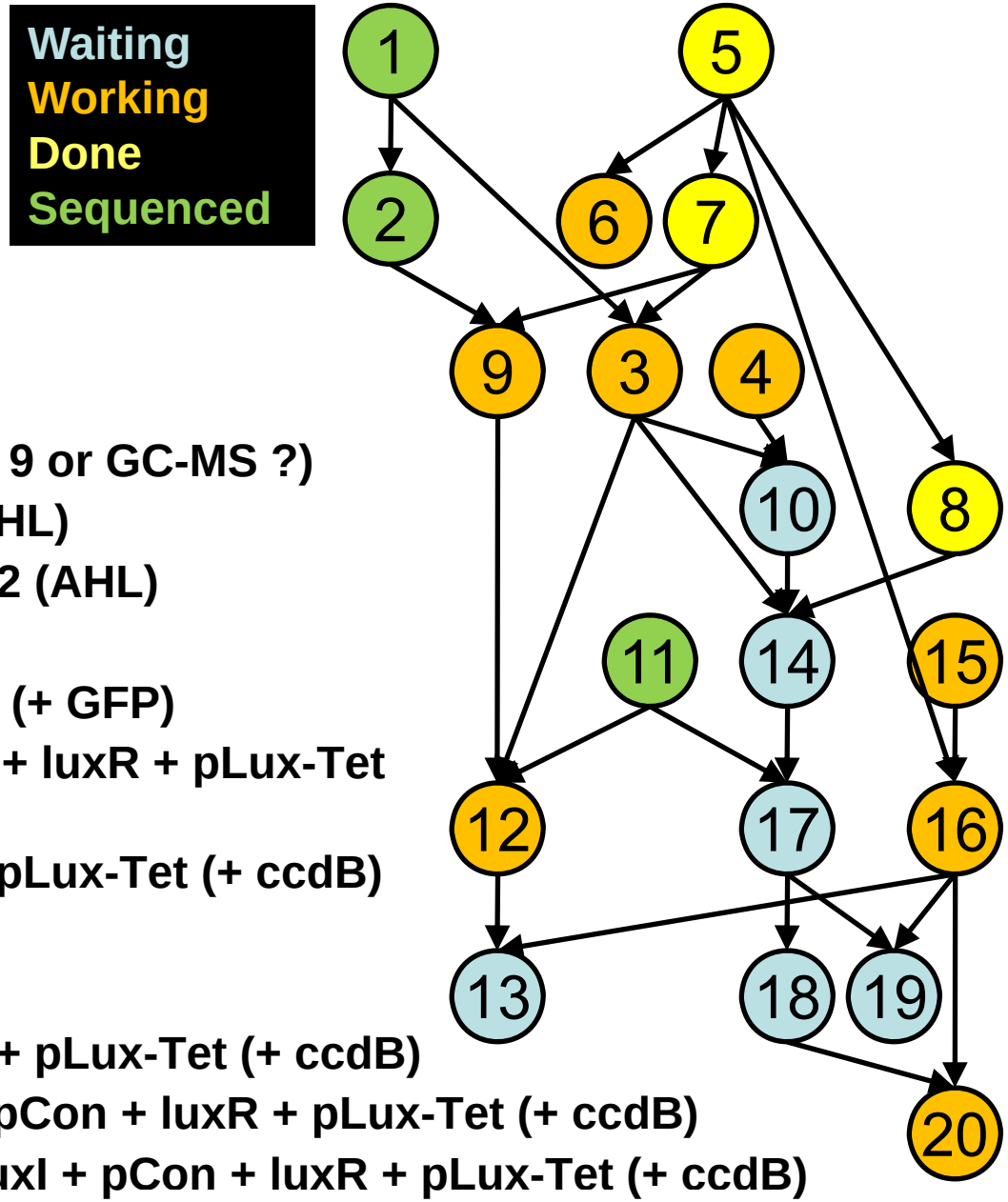
Outline

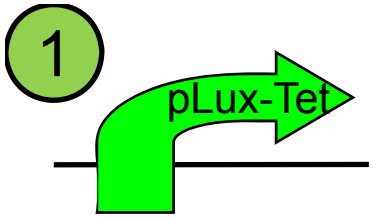
- Assembly
- Measurement
  - General Conditions
  - GFP
  - AHL
  - CcdB
  - LacZα
- Wiki
  - Team project description
  - Notebook
- Instructional Videos

Assembly

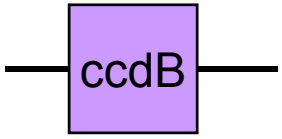

Scoring Function
Reporter
Quorum Sensing
Automatic Controller
pCon X1
VR X1
34
GFP
34
luxI-LVA
34
tetR(-LVA)
15
Quorum Sensing
pCon X2
34
luxI-LVA
15
pCon 0.70
34
luxR
15
Quorum Sensing
Selection Function
pLux-Tet
34
(lacZα-)ccdB
15

1. pLux-Tet
2. pLux-Tet + GFP
3. pCon + luxR + pLux-Tet
4. ccdB×2
5. pCon×8
6. pCon×8 + GFP
7. pCon + luxR
8. pCon×7 + luxI (AHL detection by 9 or GC-MS ?)
9. pCon + luxR + pLux-Tet + GFP (AHL)
10. pCon + luxR + pLux-Tet + ccdB×2 (AHL)
11. tetR×2
12. tetR×2 + pCon + luxR + pLux-Tet (+ GFP)
13. (VR + pCon)×8 + tetR×2 + pCon + luxR + pLux-Tet (+ GFP) (AHL/aTc)
14. (pCon×7 +) luxI + pCon + luxR + pLux-Tet (+ ccdB)
15. VR×8
16. (VR + pCon)×8
17. tetR + pCon + luxI + pCon + luxR + pLux-Tet (+ ccdB)
18. GFP + luxI + tetR + pCon + luxI + pCon + luxR + pLux-Tet (+ ccdB)
19. (VR + pCon)×8 + tetR + pCon + luxI + pCon + luxR + pLux-Tet (+ ccdB)
20. (VR + pCon)×8 + GFP + luxI + tetR + pCon + luxI + pCon + luxR + pLux-Tet (+ ccdB)



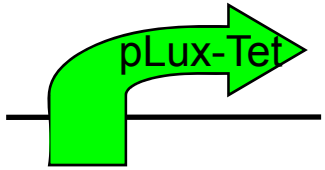


K176000 X+P with nicks
Sequence OK



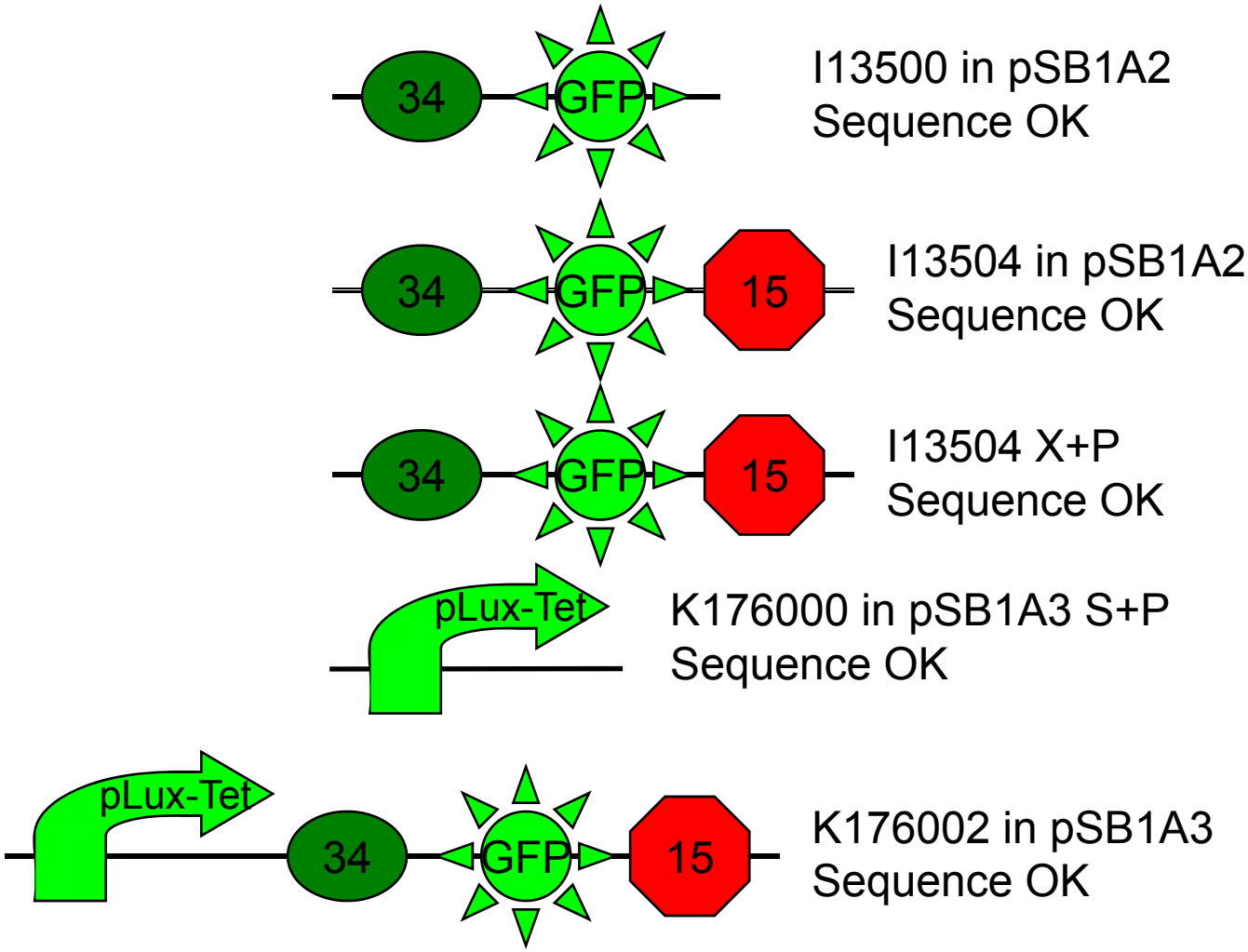
P1010 in pSB1A3 in DB 3.1
Length OK

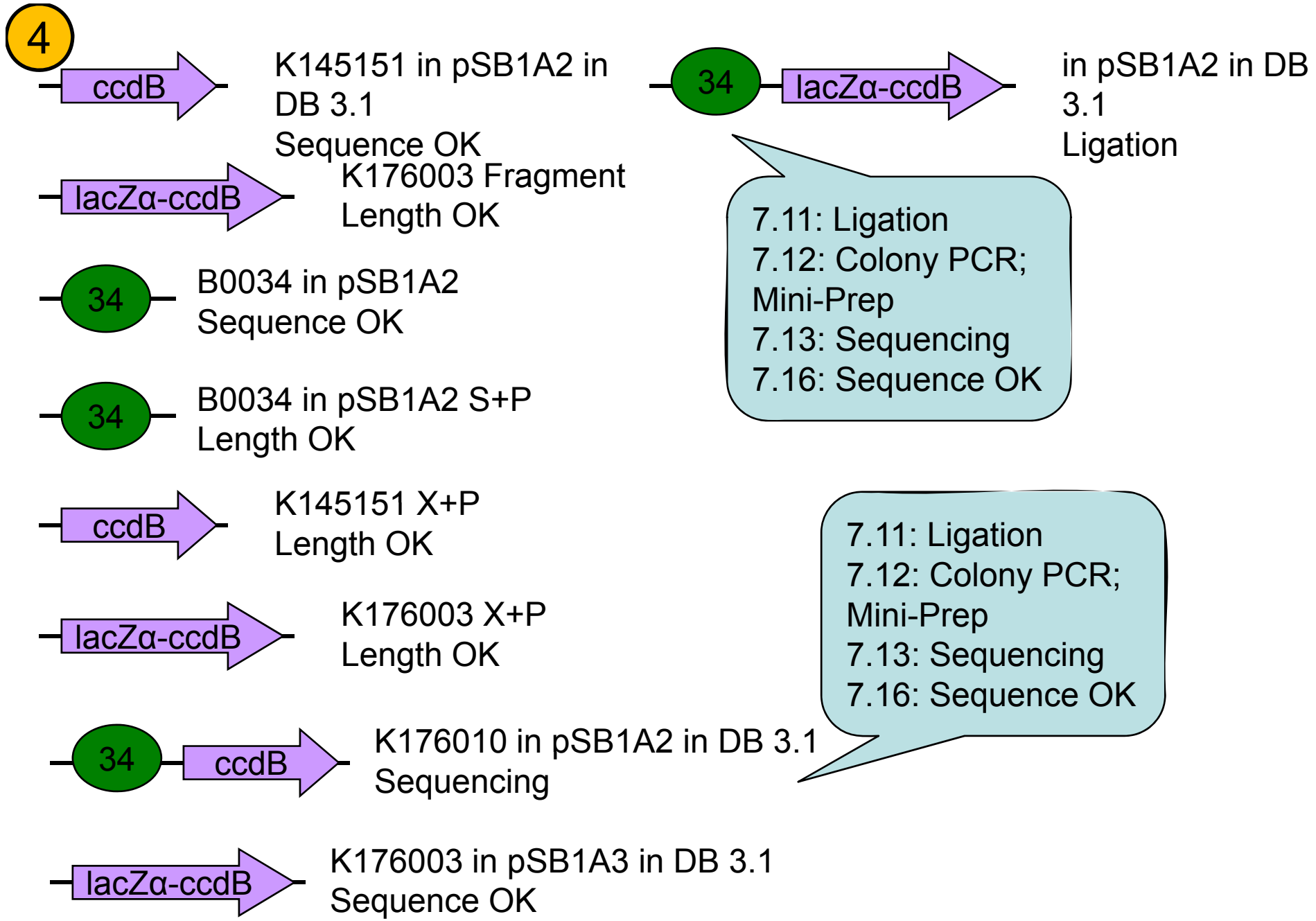
pSB1A3 X+P
Length OK

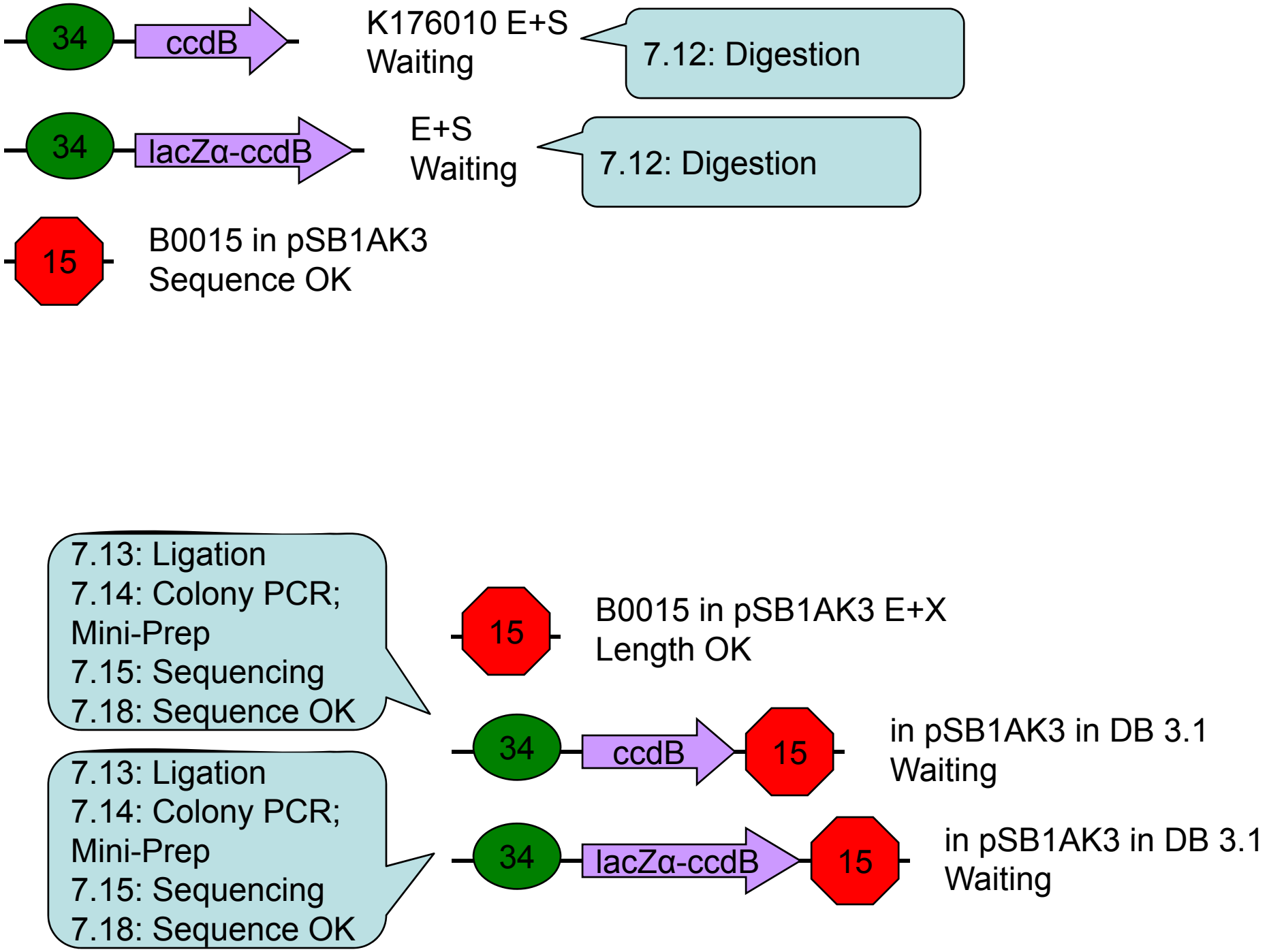


K176000 in pSB1A3
Sequence OK

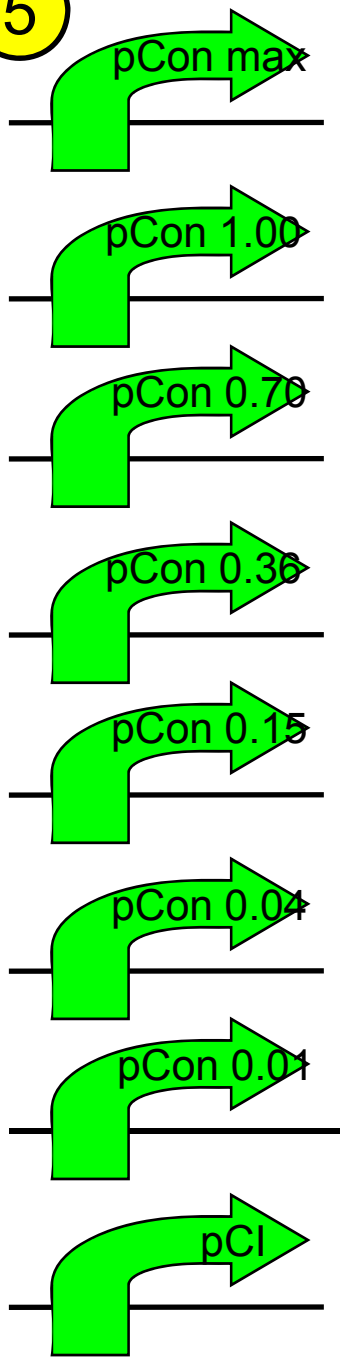
2


34
GFP
I13500 in pSB1A2
Sequence OK
34
GFP
15
I13504 in pSB1A2
Sequence OK
34
GFP
15
I13504 X+P
Sequence OK
pLux-Tet
K176000 in pSB1A3 S+P
Sequence OK
pLux-Tet
34
GFP
15
K176002 in pSB1A3
Sequence OK


4
ccdB
K145151 in pSB1A2 in DB 3.1
Sequence OK
lacZα-ccdB
K176003 Fragment
Length OK
34
B0034 in pSB1A2
Sequence OK
34
B0034 in pSB1A2 S+P
Length OK
ccdB
K145151 X+P
Length OK
lacZα-ccdB
K176003 X+P
Length OK
34
ccdB
K176010 in pSB1A2 in DB 3.1
Sequencing
lacZα-ccdB
K176003 in pSB1A3 in DB 3.1
Sequence OK
34
lacZα-ccdB
in pSB1A2 in DB 3.1
Ligation
7.11: Ligation
7.12: Colony PCR; Mini-Prep
7.13: Sequencing
7.16: Sequence OK
7.11: Ligation
7.12: Colony PCR; Mini-Prep
7.13: Sequencing
7.16: Sequence OK


34
ccdB
K176010 E+S
Waiting
7.12: Digestion
34
lacZα-ccdB
E+S
Waiting
7.12: Digestion
15
B0015 in pSB1AK3
Sequence OK
7.13: Ligation
7.14: Colony PCR;
Mini-Prep
7.15: Sequencing
7.18: Sequence OK
15
B0015 in pSB1AK3 E+X
Length OK
34
ccdB
15
in pSB1AK3 in DB 3.1
Waiting
7.13: Ligation
7.14: Colony PCR;
Mini-Prep
7.15: Sequencing
7.18: Sequence OK
34
lacZα-ccdB
15
in pSB1AK3 in DB 3.1
Waiting

5



J23119 in pSB1A2
Sequence OK

J23100 in J61002
Sequence ?

J23101 in J61002
Sequence OK

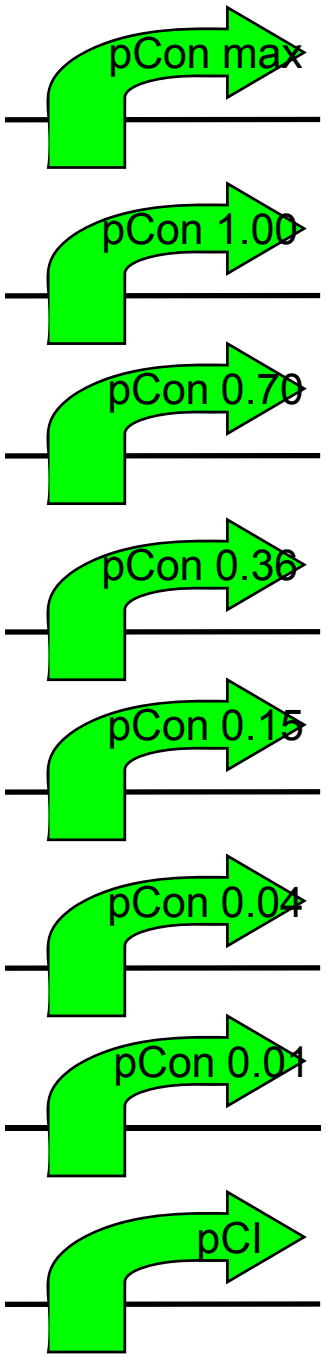
K176009 in J61002
Sequence OK

K176008 in J61002
Sequence OK

J23109 in J61002
Sequence OK

J23103 in J61002
Sequence OK

R0051 in pSB1A2
Sequence ?



J23119 X+P
Sequence OK

J23100-J61002SF X+P
Sequence ?

J23101-J61002SF X+P
Sequence OK

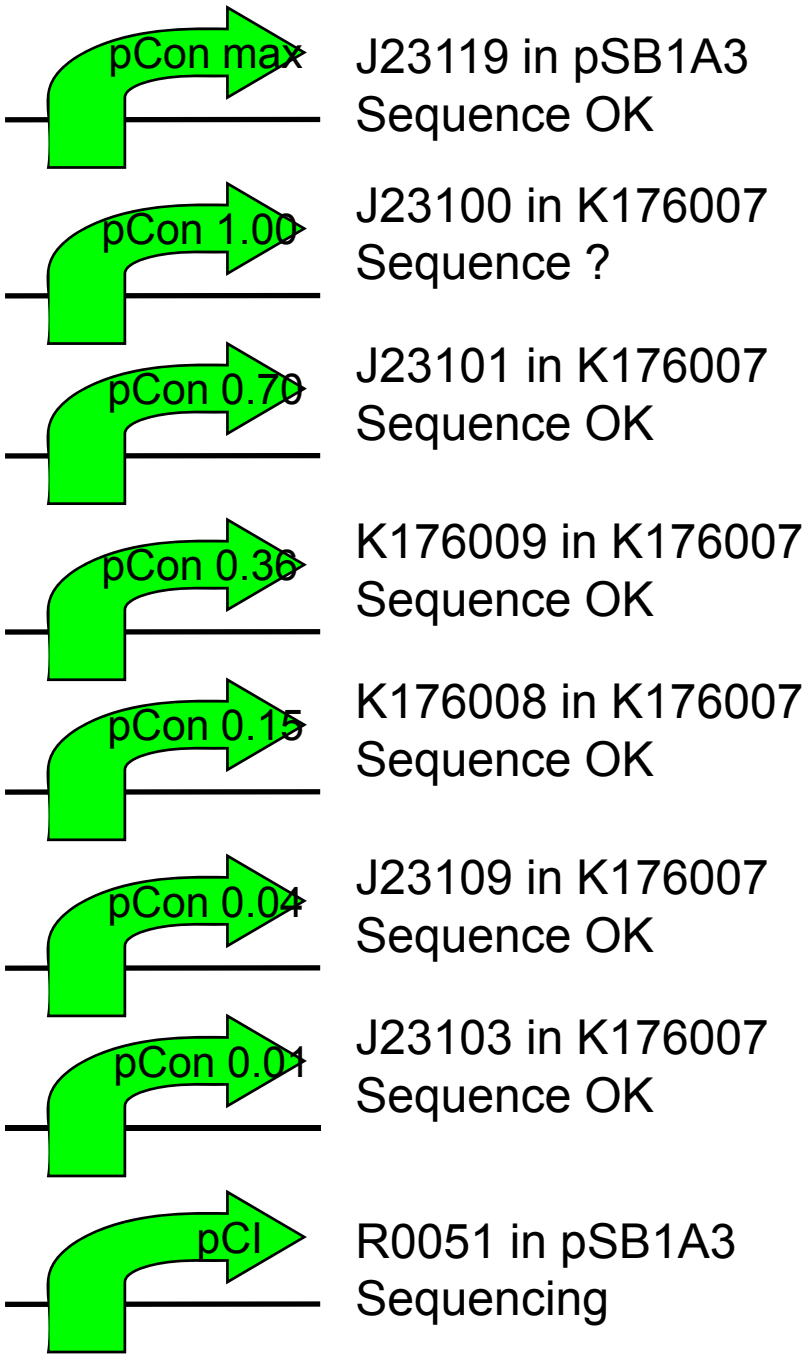
K176009-J61002SF X+P
Sequence OK

K176008-J61002SF X+P
Sequence OK

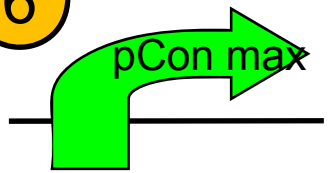
J23109-J61002SF X+P
Sequence OK

J23103-J61002SF X+P
Sequence OK

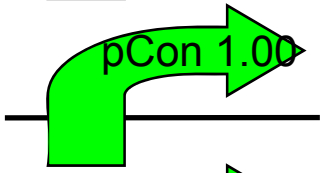
R0051 X+P
Length ?


pCon max
J23119 in pSB1A3
Sequence OK
pCon 1.00
J23100 in K176007
Sequence ?
pCon 0.70
J23101 in K176007
Sequence OK
pCon 0.36
K176009 in K176007
Sequence OK
pCon 0.15
K176008 in K176007
Sequence OK
pCon 0.04
J23109 in K176007
Sequence OK
pCon 0.01
J23103 in K176007
Sequence OK
pCI
R0051 in pSB1A3
Sequencing

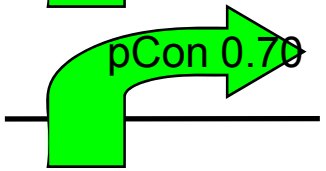
6



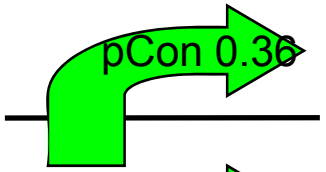
J23119 in pSB1A3 S+P
Length OK



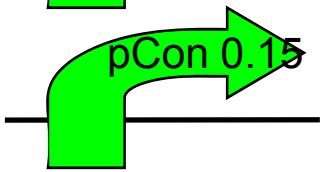
J23100 in K176007 S+P
Digestion



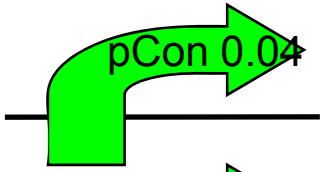
J23101 in K176007 S+P
Length OK



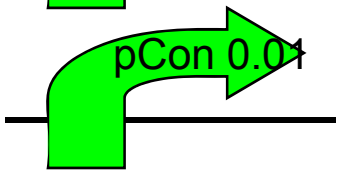
K176009 in K176007 S+P
Length OK



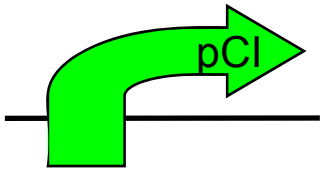
K176008 in K176007 S+P
Length OK



J23109 in K176007 S+P
Length OK

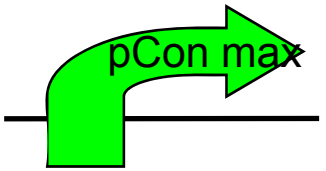


J23103 in K176007 S+P
Length OK

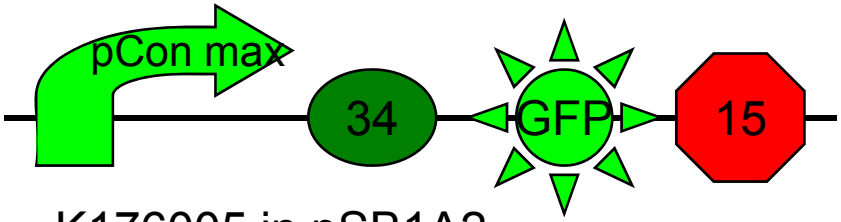


R0051 in pSB1A3 S+P
Digestion

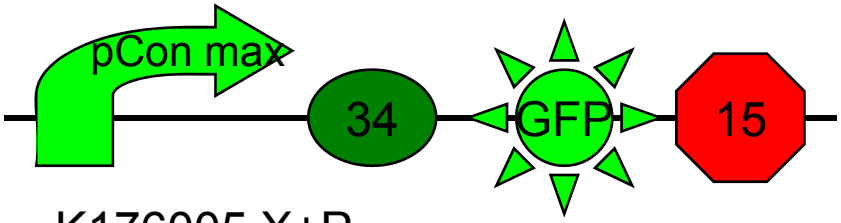
7.12: Digestion



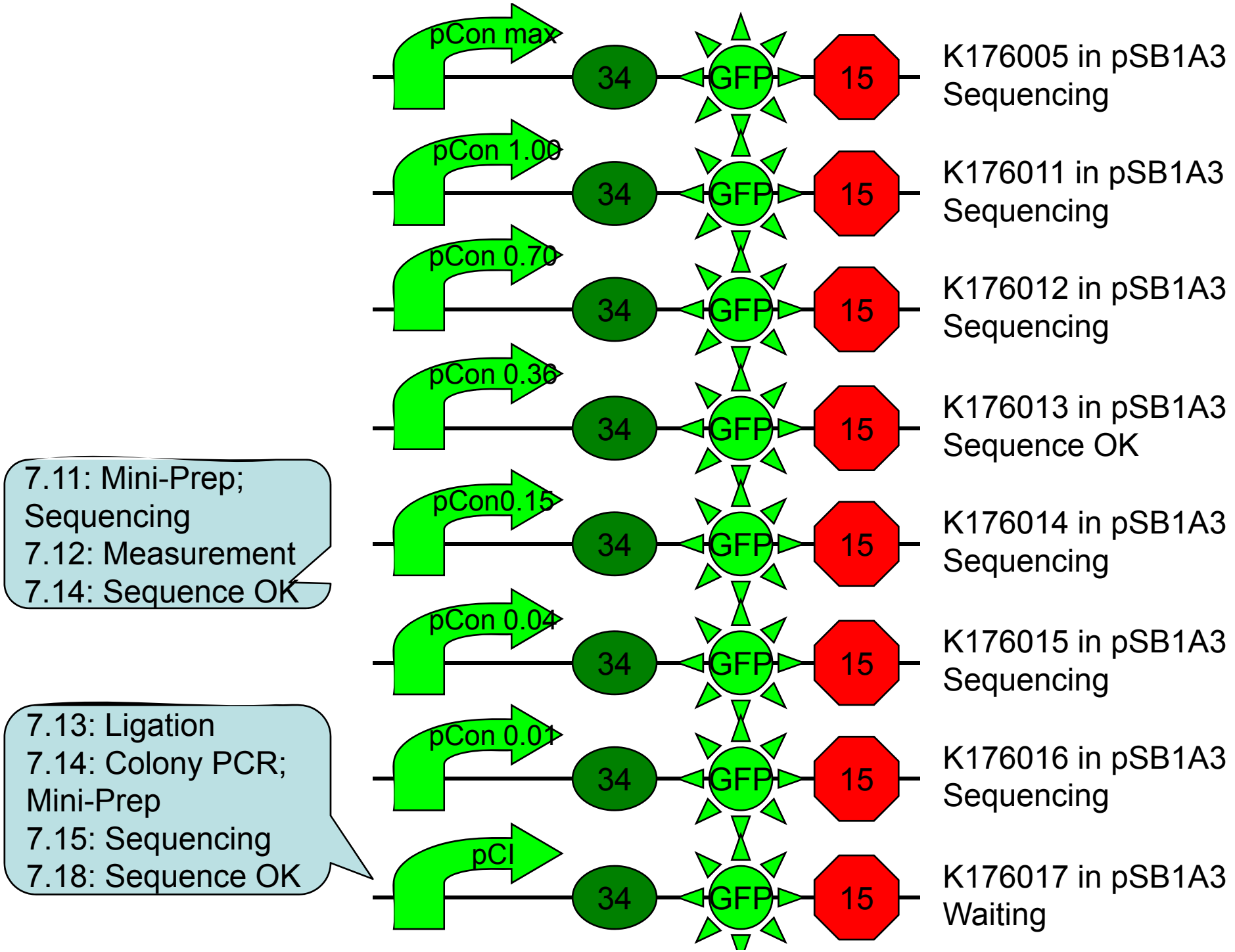
J23119 in pSB1A2 S+P
Sequence OK

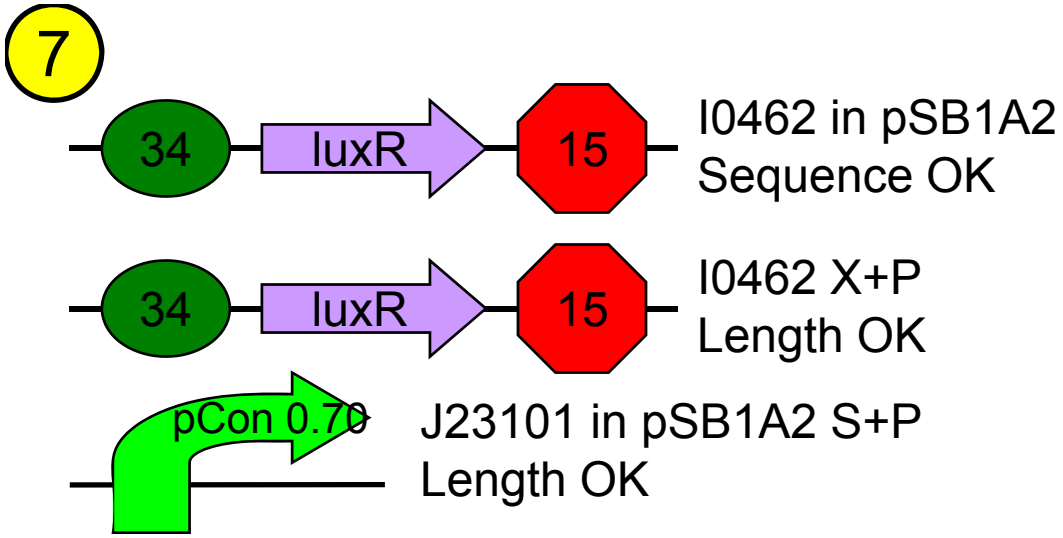


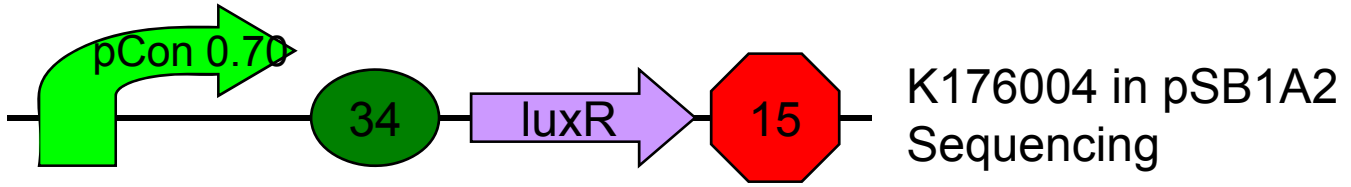
K176005 in pSB1A2
Sequence OK

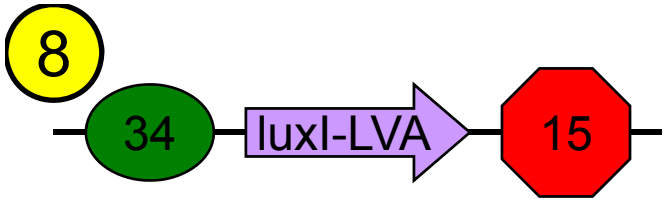


K176005 X+P
Length OK


pCon max
34
GFP
15
K176005 in pSB1A3
Sequencing
pCon 1.00
34
GFP
15
K176011 in pSB1A3
Sequencing
pCon 0.70
34
GFP
15
K176012 in pSB1A3
Sequencing
pCon 0.36
34
GFP
15
K176013 in pSB1A3
Sequence OK
pCon0.15
34
GFP
15
K176014 in pSB1A3
Sequencing
pCon 0.04
34
GFP
15
K176015 in pSB1A3
Sequencing
pCon 0.01
34
GFP
15
K176016 in pSB1A3
Sequencing
pCl
34
GFP
15
K176017 in pSB1A3
Waiting
7.11: Mini-Prep;
Sequencing
7.12: Measurement
7.14: Sequence OK
7.13: Ligation
7.14: Colony PCR;
Mini-Prep
7.15: Sequencing
7.18: Sequence OK


7
34
luxR
15
I0462 in pSB1A2
Sequence OK
34
luxR
15
I0462 X+P
Length OK
pCon 0.70
J23101 in pSB1A2 S+P
Length OK

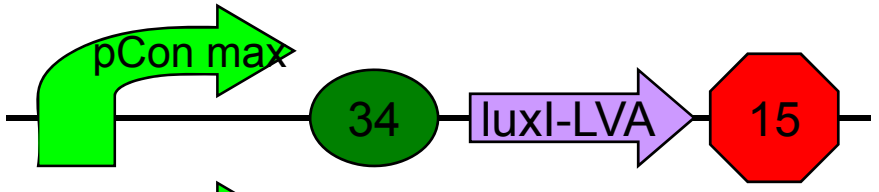

pCon 0.70
34
luxR
15
K176004 in pSB1A2
Sequencing



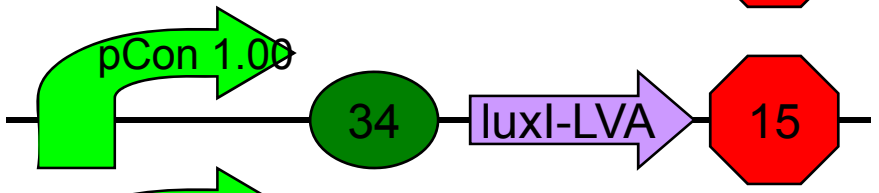
K082014 X+P
Length OK

7.11: Mini-Prep;
Sequencing
7.12: Measurement
7.14: Sequence OK

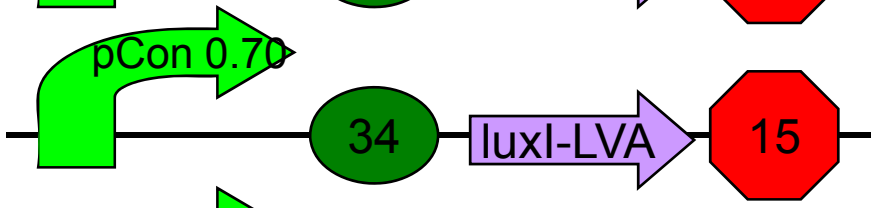
K082014 in pSB1AK3
Sequence OK



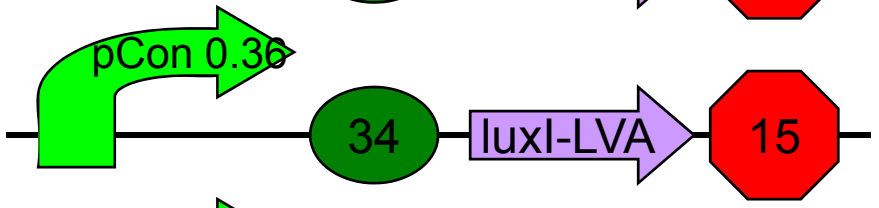
K176018 in
pSB1A3
Sequencing



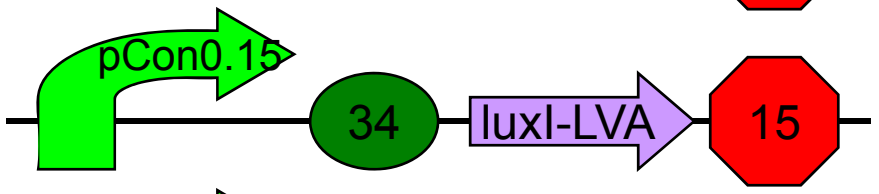
K176019 in
pSB1A3
Sequencing



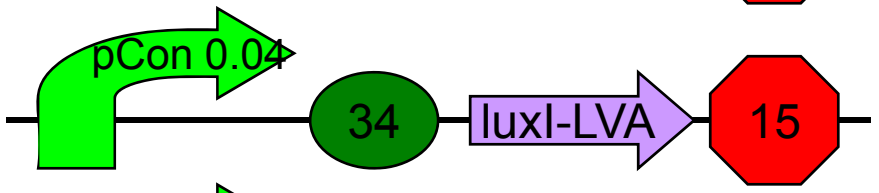
K176020 in
pSB1A3
Sequencing



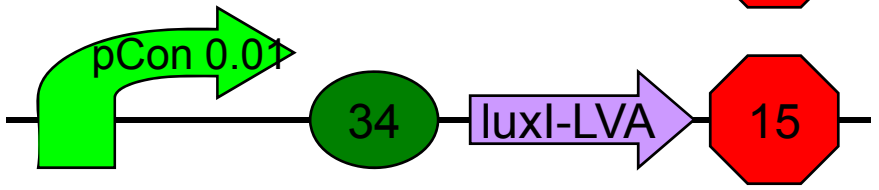
K176021 in
pSB1A3
Sequencing



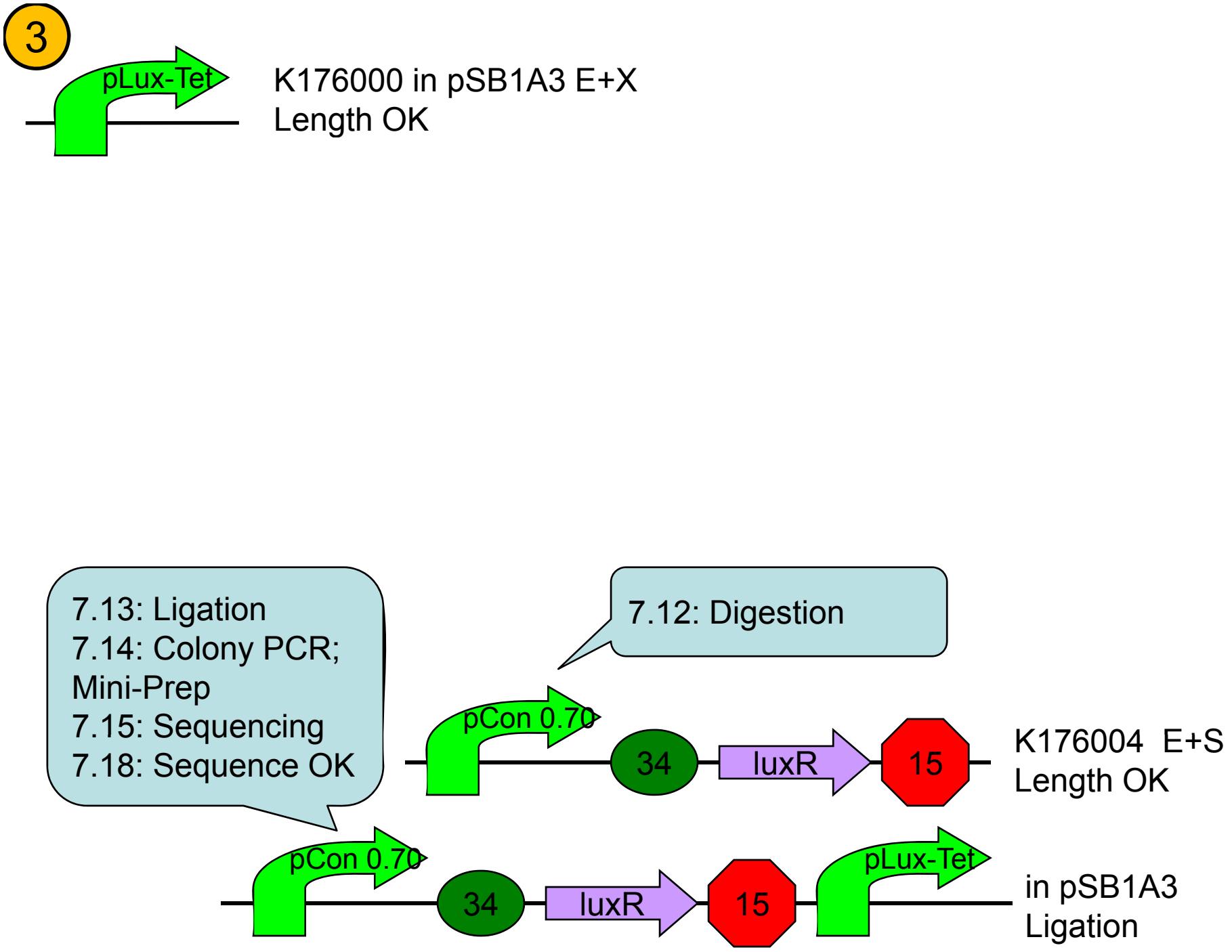
K176022 in
pSB1A3
Sequencing

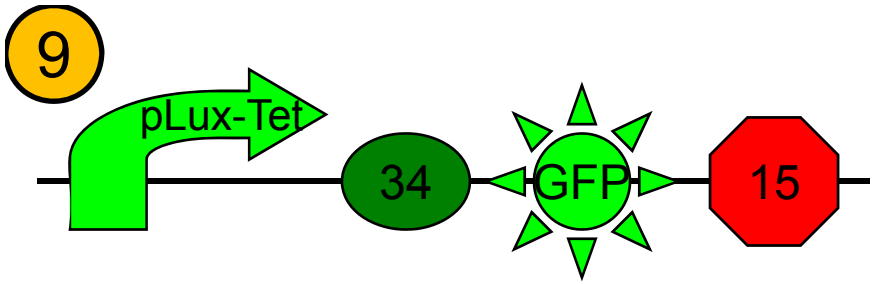


K176023 in
pSB1A3
Sequencing



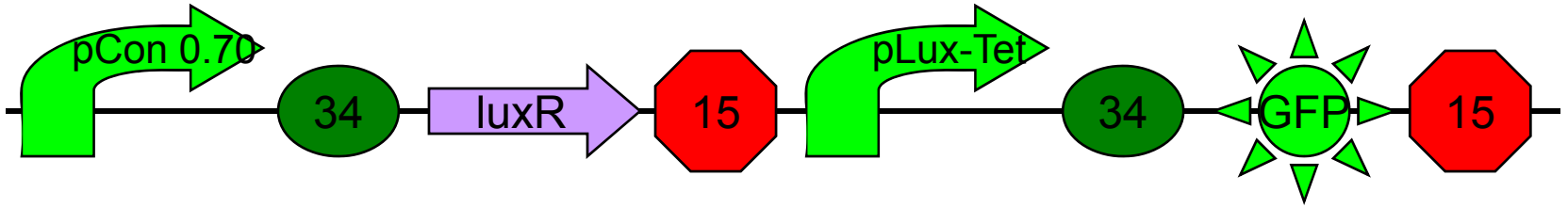
K176024 in
pSB1A3
Sequencing


3
pLux-Tet
K176000 in pSB1A3 E+X
Length OK
7.13: Ligation
7.14: Colony PCR;
Mini-Prep
7.15: Sequencing
7.18: Sequence OK
7.12: Digestion
pCon 0.70
34
luxR
15
K176004 E+S
Length OK
pCon 0.70
34
luxR
15
pLux-Tet
in pSB1A3
Ligation

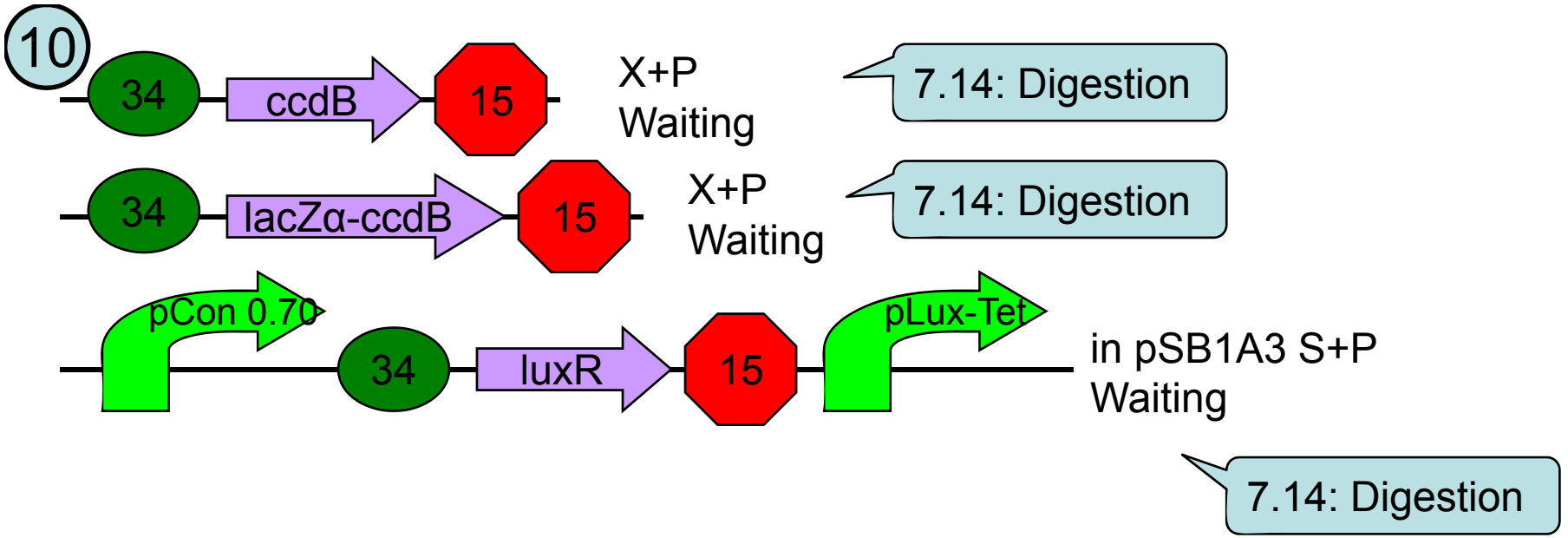


K176002 in pSB1A3 E+X
Length OK

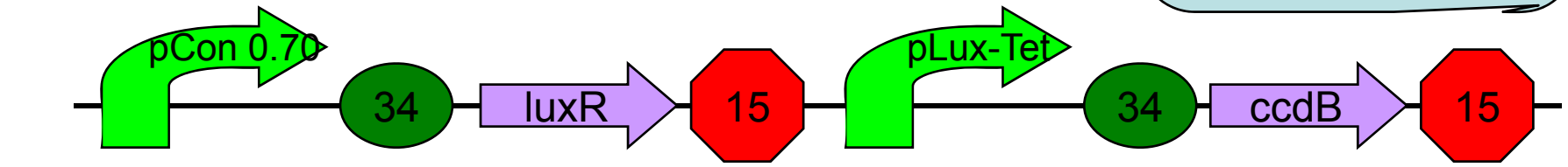
7.13: Ligation
7.14: Colony PCR;
Mini-Prep
7.15: Sequencing
7.16: Measurement
7.18: Sequence OK



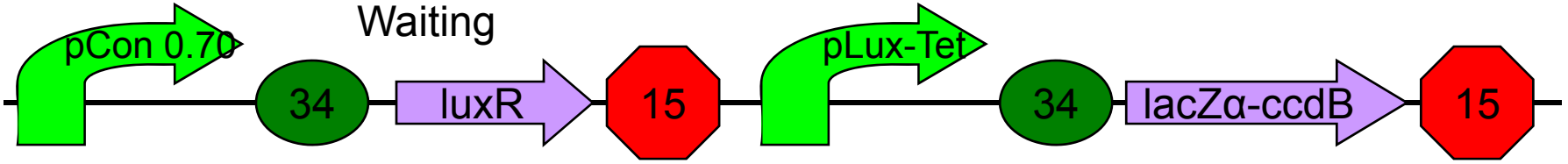
in pSB1A3
Ligation


10
34
ccdB
15
X+P
Waiting
7.14: Digestion
34
lacZα-ccdB
15
X+P
Waiting
7.14: Digestion
pCon 0.70
34
luxR
15
pLux-Tet
in pSB1A3 S+P
Waiting
7.14: Digestion

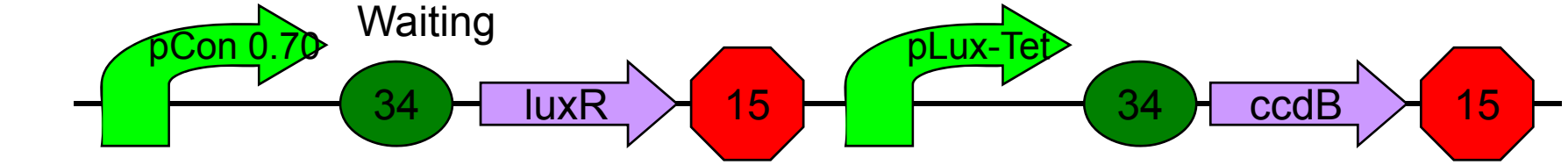
7.15: Ligation
7.16: Colony PCR; Mini-Prep
7.17: Sequencing
7.18: Measurement
7.20: Sequence OK



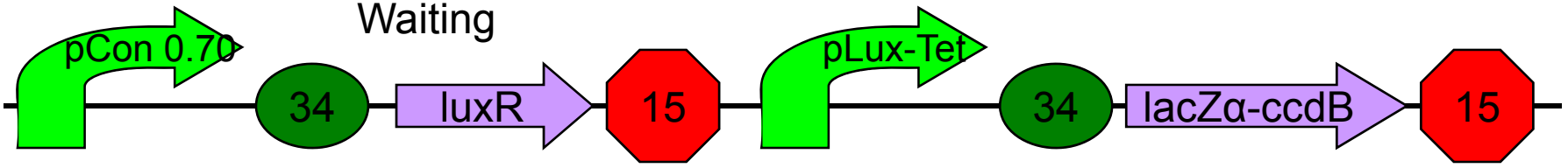
in pSB1A3 in DB 3.1
Waiting



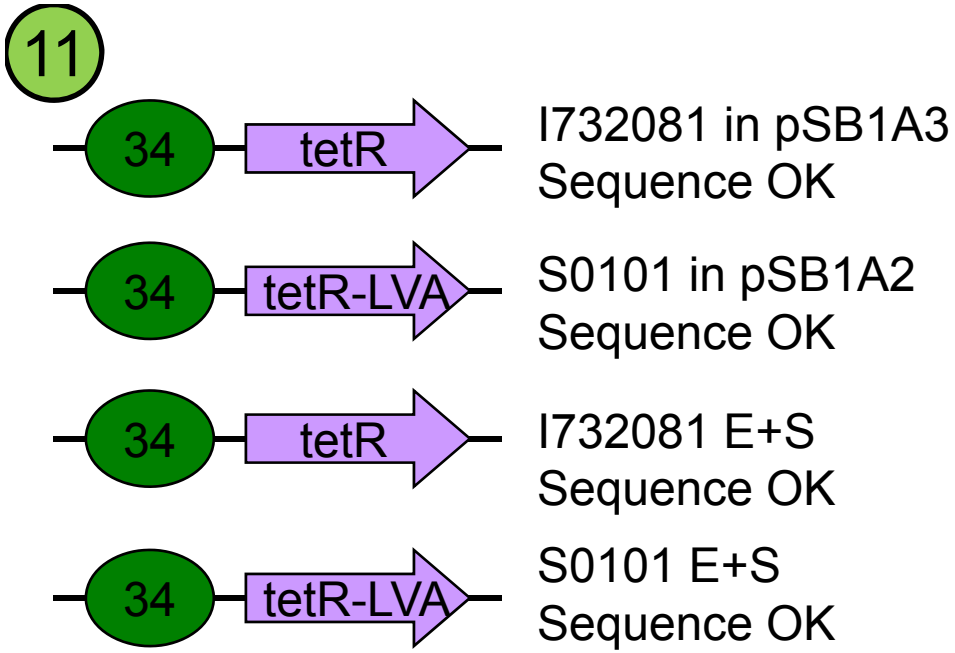
in pSB1A3 in DB 3.1
Waiting

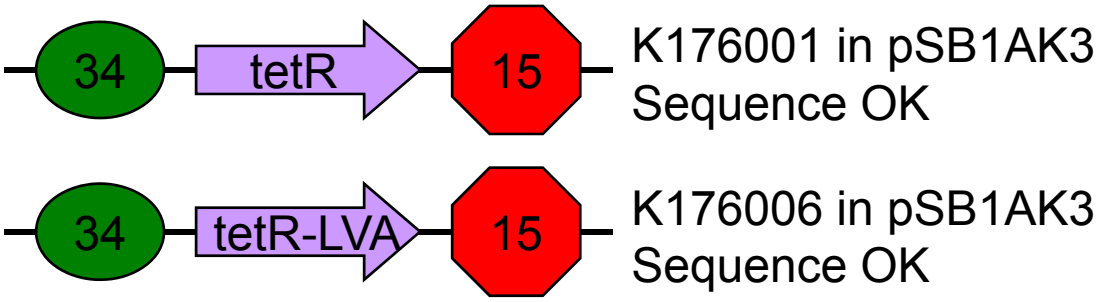


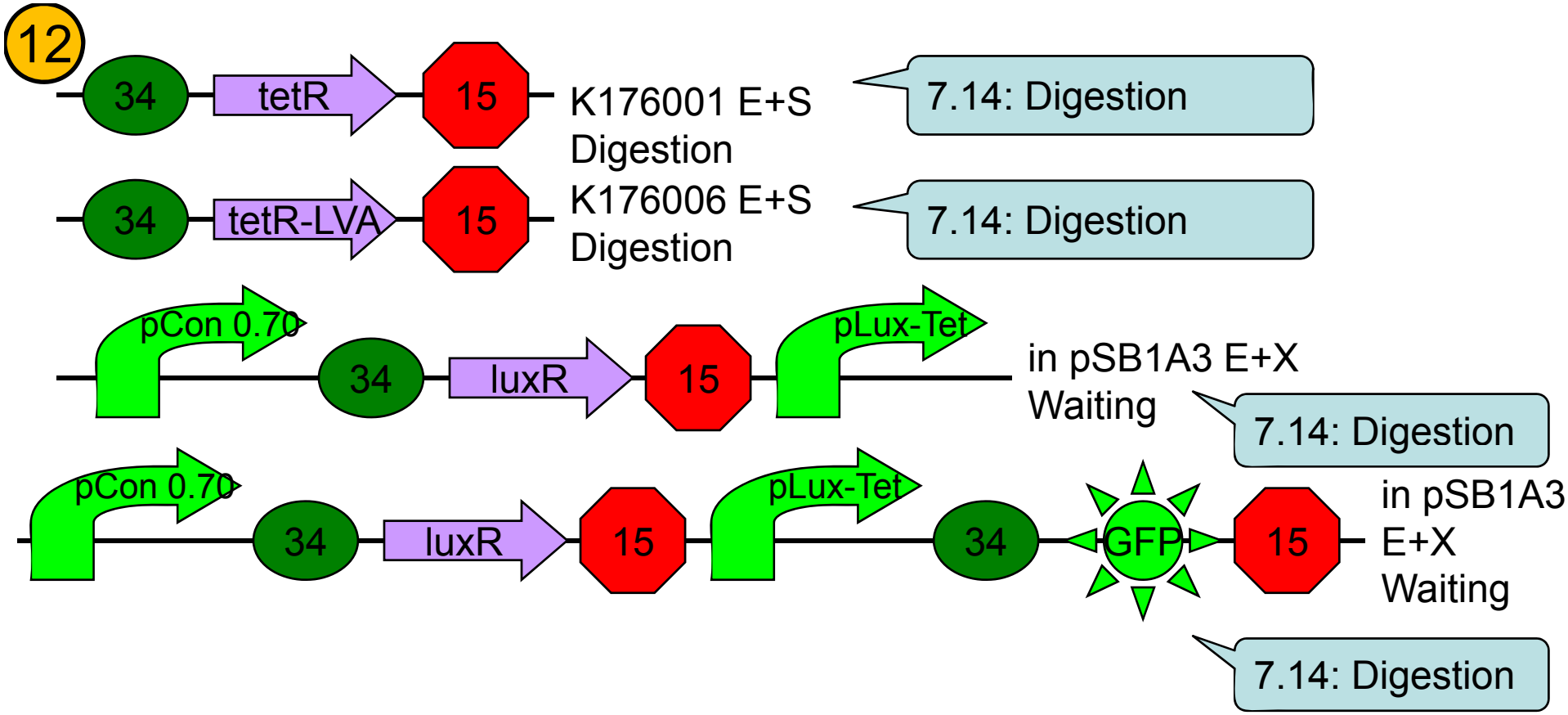
in pSB1A3 in TOP10
Waiting



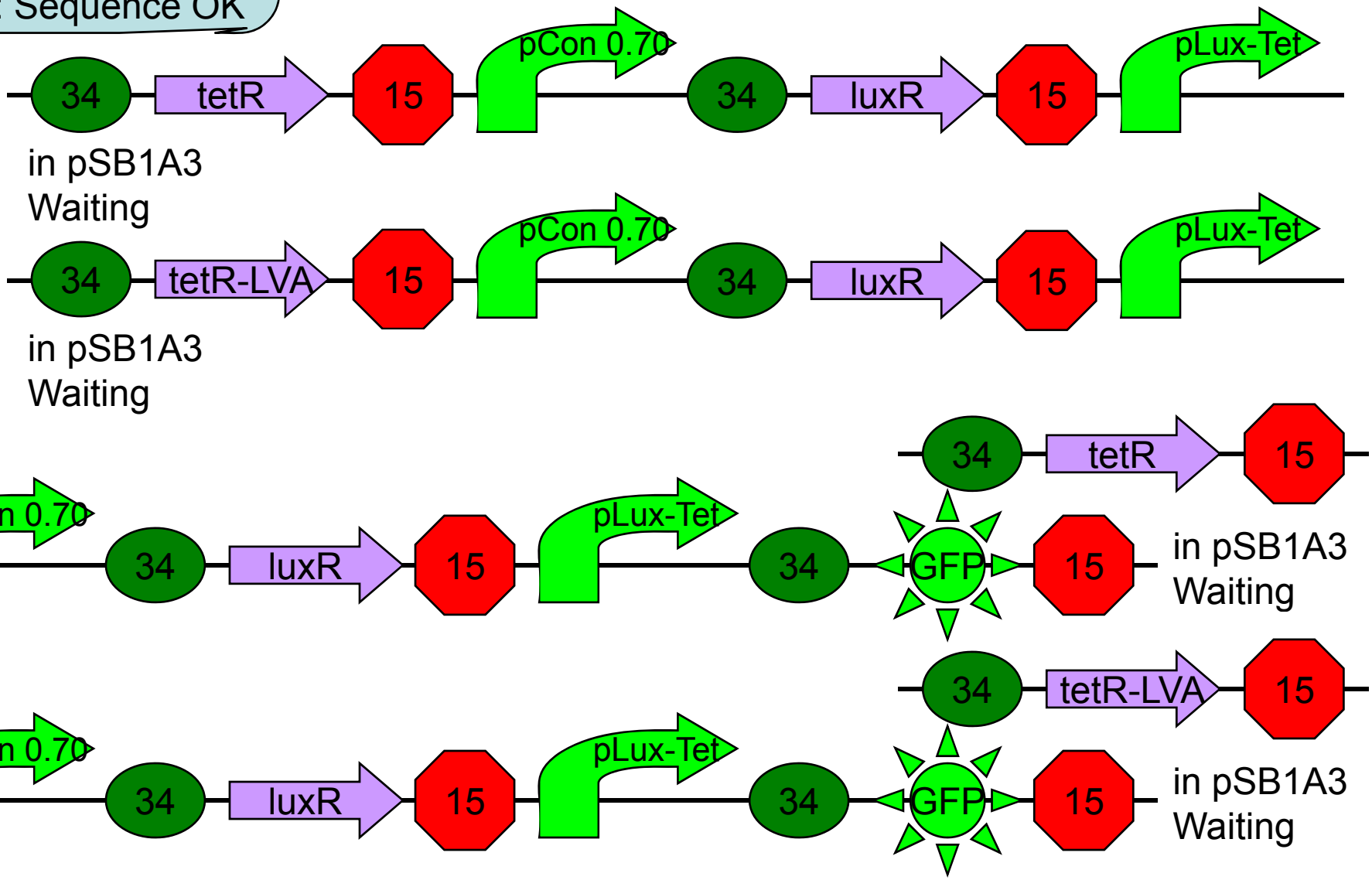
in pSB1A3 in TOP10
Waiting

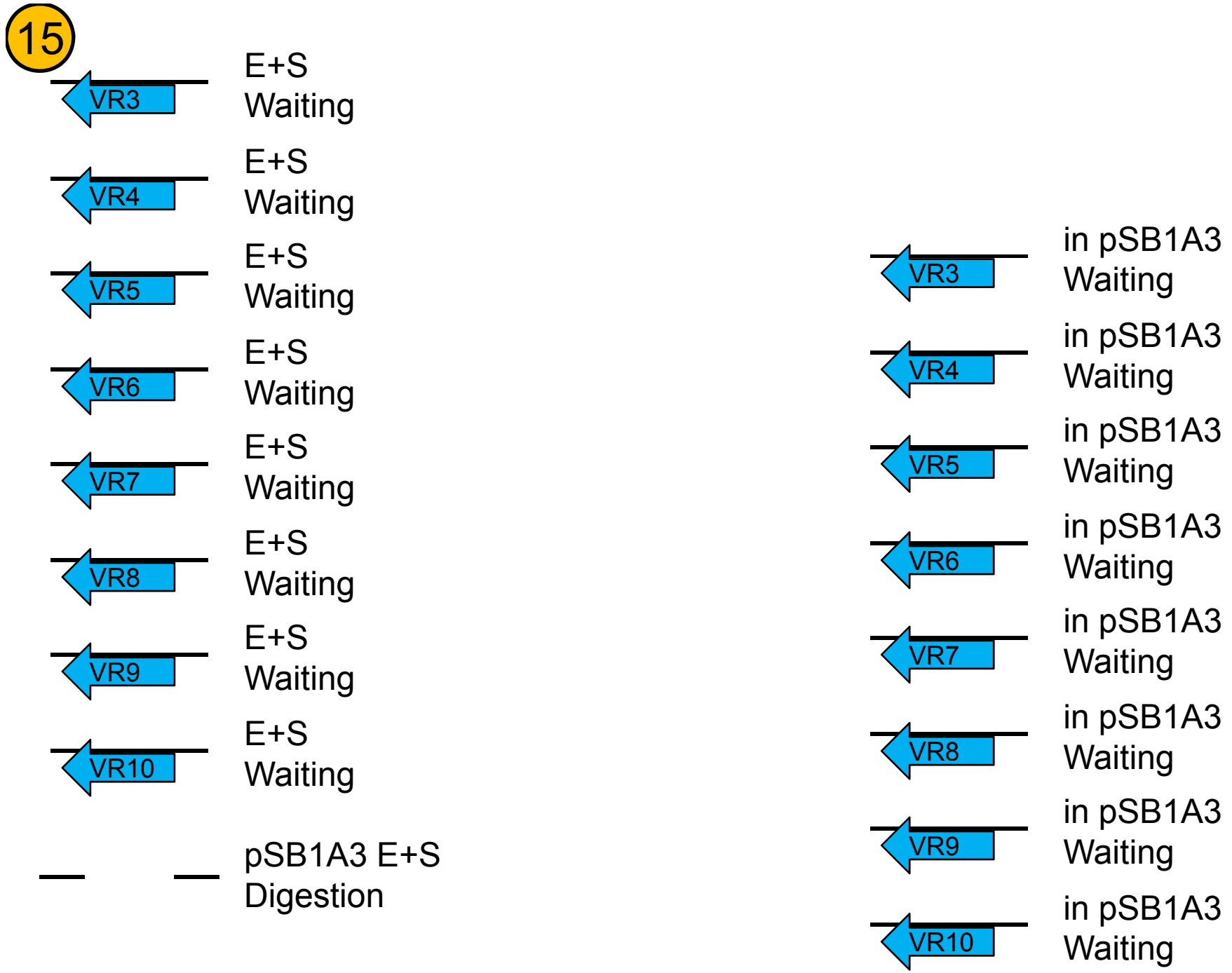

11
34
tetR
I732081 in pSB1A3
Sequence OK
34
tetR-LVA
S0101 in pSB1A2
Sequence OK
34
tetR
I732081 E+S
Sequence OK
34
tetR-LVA
S0101 E+S
Sequence OK


34
tetR
15
K176001 in pSB1AK3
Sequence OK
34
tetR-LVA
15
K176006 in pSB1AK3
Sequence OK

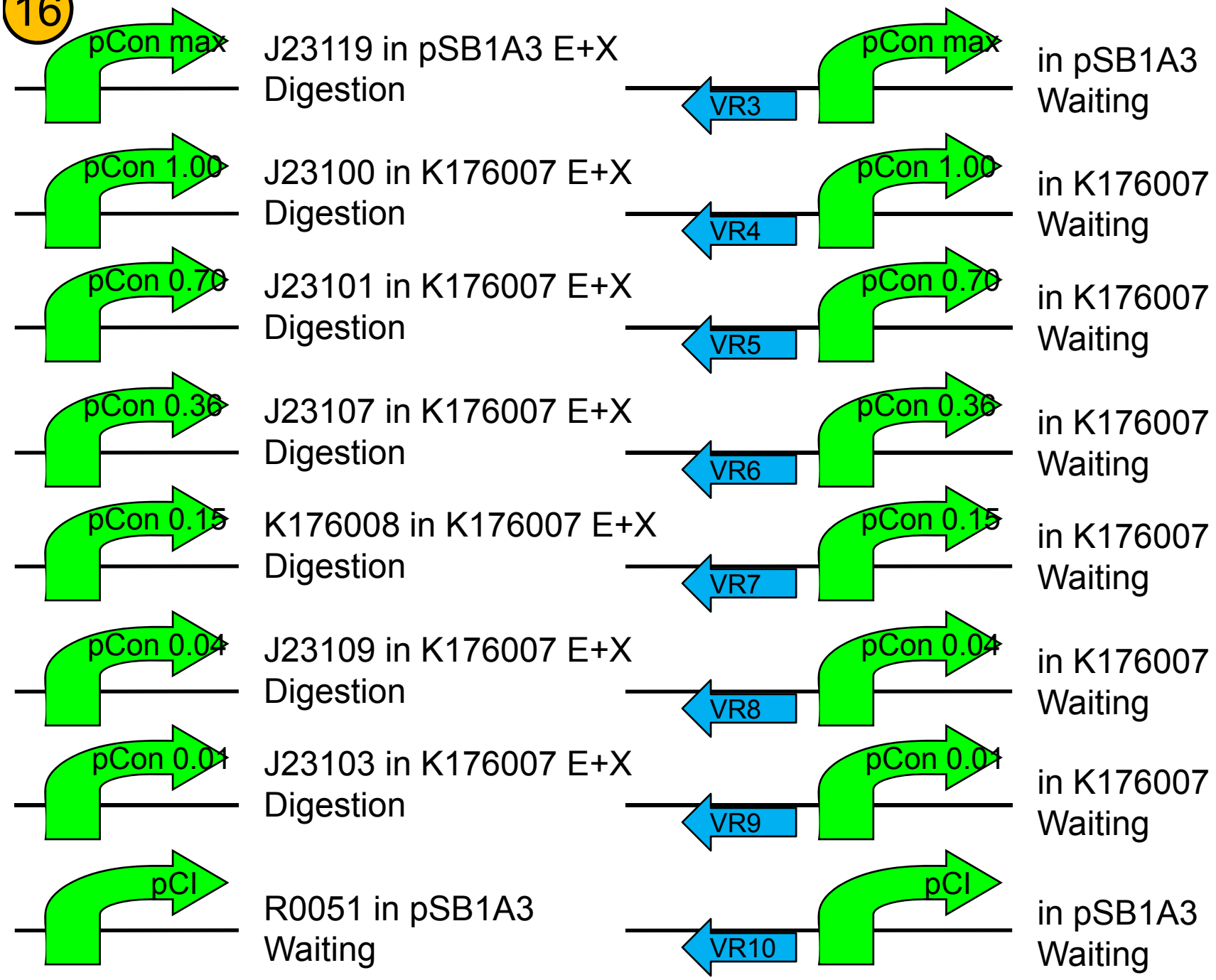

12
34
tetR
15
K176001 E+S
Digestion
7.14: Digestion
34
tetR-LVA
15
K176006 E+S
Digestion
7.14: Digestion
pCon 0.70
34
luxR
15
pLux-Tet
in pSB1A3 E+X
Waiting
7.14: Digestion
pCon 0.70
34
luxR
15
pLux-Tet
34
GFP
15
in pSB1A3
E+X
Waiting
7.14: Digestion

7.15: Ligation
7.16: Colony PCR;
Mini-Prep
7.17: Sequencing
7.20: Sequence OK




15
VR3
E+S
Waiting
VR4
E+S
Waiting
VR5
E+S
Waiting
VR6
E+S
Waiting
VR7
E+S
Waiting
VR8
E+S
Waiting
VR9
E+S
Waiting
VR10
E+S
Waiting
pSB1A3 E+S
Digestion
VR3
in pSB1A3
Waiting
VR4
in pSB1A3
Waiting
VR5
in pSB1A3
Waiting
VR6
in pSB1A3
Waiting
VR7
in pSB1A3
Waiting
VR8
in pSB1A3
Waiting
VR9
in pSB1A3
Waiting
VR10
in pSB1A3
Waiting

16

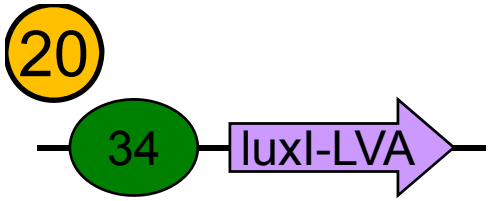

pCon max
J23119 in pSB1A3 E+X
Digestion
VR3
pCon max
in pSB1A3
Waiting
pCon 1.00
J23100 in K176007 E+X
Digestion
VR4
pCon 1.00
in K176007
Waiting
pCon 0.70
J23101 in K176007 E+X
Digestion
VR5
pCon 0.70
in K176007
Waiting
pCon 0.36
J23107 in K176007 E+X
Digestion
VR6
pCon 0.36
in K176007
Waiting
pCon 0.15
K176008 in K176007 E+X
Digestion
VR7
pCon 0.15
in K176007
Waiting
pCon 0.04
J23109 in K176007 E+X
Digestion
VR8
pCon 0.04
in K176007
Waiting
pCon 0.01
J23103 in K176007 E+X
Digestion
VR9
pCon 0.01
in K176007
Waiting
pCI
R0051 in pSB1A3
Waiting
VR10
pCI
in pSB1A3
Waiting

13

14

17

18



C0261 in pSB1A2
Sequence OK

Favorite USTC 2009 iGEM Team Parts

Edit

-?-	Name	Type	Description	Designer	Length

USTC 2009 iGEM Team Parts Sandbox

Edit

-?-		Name	Type	Description	Designer	Length
		BBa_K176000	Regulatory	pLux/Tet Hybrid Promoter: (LuxR+,TetR-)->PoPS	Danqian Liu, Chao Li, Hao Jiang	72
		BBa_K176001	Generator	PoPS->RBS+TetR(without LVA)+T	Chao Li,Danqian Liu,Hao Jiang	782
		BBa_K176002	Reporter	pLux/Tet(K176000)(LuxR+,TetR-)->RBS+GFP+T	Chao Li,Danqian Liu,Hao Jiang	955
		BBa_K176003	Coding	lacZalpha-ccdB coding sequence	Zongxiao He, Hao Jiang	480
		BBa_K176004	Generator	pCon max(J23119)->RBS+LuxR+T	Chao Li,Danqian Liu,Hao Jiang	979
		BBa_K176005	Reporter	pCon max(J23119)->RBS+GFP+T	Chao Li,Danqian Liu,Hao Jiang	918
		BBa_K176006	Generator	PoPS->RBS+TetR(with LVA)+T	Chao Li,Danqian Liu,Hao Jiang	840
	W	BBa_K176007	Plasmid_Backbone	pSB1A3 with the suffix of J61002 (mRFP)	Hao Jiang, Danqian Liu, Chao Li	3026
		BBa_K176008	Regulatory	constitutive promoter family member J23115 actual sequence	Hao Jiang, Danqian Liu, Chao Li	35
		BBa_K176009	Regulatory	constitutive promoter family member J23107 actual sequence	Hao Jiang, Danqian Liu, Chao Li	35
		BBa_K176010	Translational_Unit	PoPS->RBS+ccdB->PoPS	Zongxiao He, Hao Jiang	324
		BBa_K176011	Reporter	pCon 1.00(J23100)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
		BBa_K176012	Reporter	pCon 0.70(J23101)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
		BBa_K176013	Reporter	pCon 0.36(K176009)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
		BBa_K176014	Reporter	pCon 0.15(K176008)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
		BBa_K176015	Reporter	pCon 0.04(J23109)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
		BBa_K176016	Reporter	pCon 0.01(J23103)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	918
		BBa_K176017	Reporter	pCI(R0051)(lambda CI-)->RBS+GFP+T	Chao Li, Danqian Liu, Hao Jiang	932
		BBa_K176018	Signalling	pCon max(J23119)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
		BBa_K176019	Signalling	pCon 1.00(J23100)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
		BBa_K176020	Signalling	pCon 0.70(J23101)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
		BBa_K176021	Signalling	pCon 0.36(K176009)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
		BBa_K176022	Signalling	pCon 0.15(K176008)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
		BBa_K176023	Signalling	pCon 0.04(J23109)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841
		BBa_K176024	Signalling	pCon 0.01(J23103)->RBS+LuxI-LVA+T	Danqian Liu, Chao Li, Hao Jiang	841

Measurement

- General Conditions
  - Medium
    - LB
    - M9
      - Minimal
      - Supplemented
    - EZ Rich Define
    - pH-buffered TBK
    - pH-buffered LBK
  - Temperature
    - 37℃
    - 30℃
    - 34℃
  - Pre-warm
  - Shake
  - Dilution
  - Wash

http://openwetware.org/wiki/M9_medium
http://openwetware.org/wiki/M9_medium/minimal
http://openwetware.org/wiki/M9_medium/supplemented
http://openwetware.org/wiki/Neidhardt_EZ_Rich_Defined
http://www.genome.wisc.edu/resources/protocols/ezmedium.htm

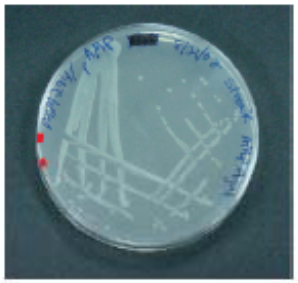
GFP (PoPS)

- Fluorospectrophotometer
- Plate Reader
- Flow Cytometer (FACS)
- Blotting
  - Northern
  - Western
- Realtime RT-PCR

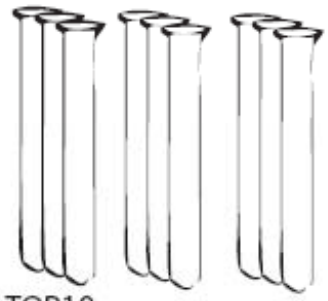
- http://partsregistry.org/Measurement
- http://openwetware.org/wiki/The_BioBricks_Foundation:Standards/Technical/Measurement
- http://openwetware.org/wiki/Standardized_GFP_quantification
- Engineering the interface between cellular chassis (Barry Canton PhD thesis)
- Applying engineering principles to the design and construction of transcriptional devices (Reshma P. Shetty PhD thesis)

STEP1: Streak 3 plates

A: TOP10
B: BBa_I20260
C: Your promoter!



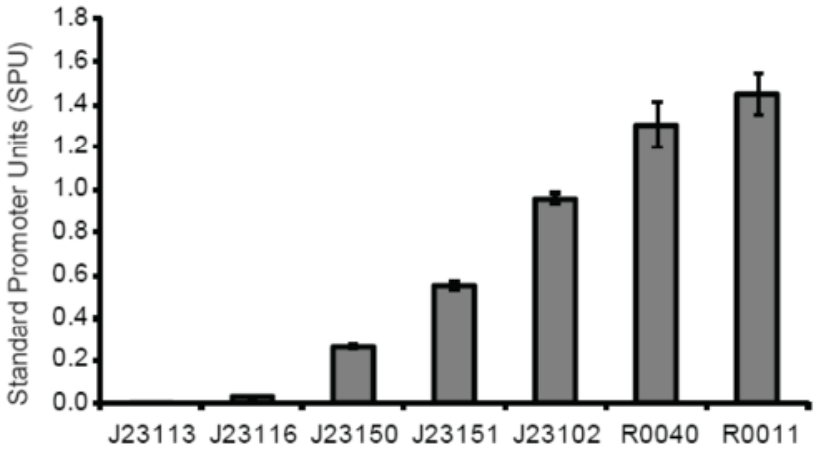
STEP 2: Pick 3 colonies from each plate to start overnight cultures in Supplemented M9 Media at 37 C (9 tubes)



TOP10
BBa_I20260
Your Promoter

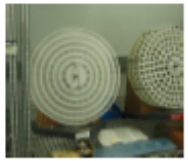
37C

16 hours



Standard Promoter Units

STEP 3: Dilute 1/100 into fresh, pre-warmed media incubate at 37C (9 tubes)



37C

STEP 4: After 3 hours measure GFP and OD

3 hours
GFP
OD

STEP 5: After another half hour measure GFP and OD again

1/2 hour
GFP
OD

2006 Berkeley **J23100~J23119** Reported activities of the promoters are given as the relative fluorescence of these plasmids in strain TG1 grown in LB media to **saturation**. See part J61002 for details on their use.

AHL

- Rapid Screening of Quorum-Sensing Signal N-Acyl Homoserine Lactones by an In Vitro Cell-Free Assay
- Detection of N-acylhomoserine lactones in lung tissues of mice infected with Pseudomonas aeruginosa
- Detecting and characterizing N-acyl-homoserine lactone signal molecules by thin-layer chromatography
- Detection of N-acyl homoserine lactones using a traI-luxCDABE-based biosensor as a high-throughput screening tool
- On-line high-performance liquid chromatography-mass spectrometric detection and quantification of N-acylhomoserine lactones, quorum sensing signal molecules, in the presence of biological matrices
- Detection of quorum-sensing N-acyl homoserine lactone signal molecules by bacterial biosensors

CcdB & LacZα

- Programmed population control by cell–cell communication and regulated killing
- A synthetic Escherichia coli predator–prey ecosystem
- LacZα
  - X-gal
  - ONPG
    - http://parts.mit.edu/igem07/index.php/USTC/BetaGalactosidaseAssay

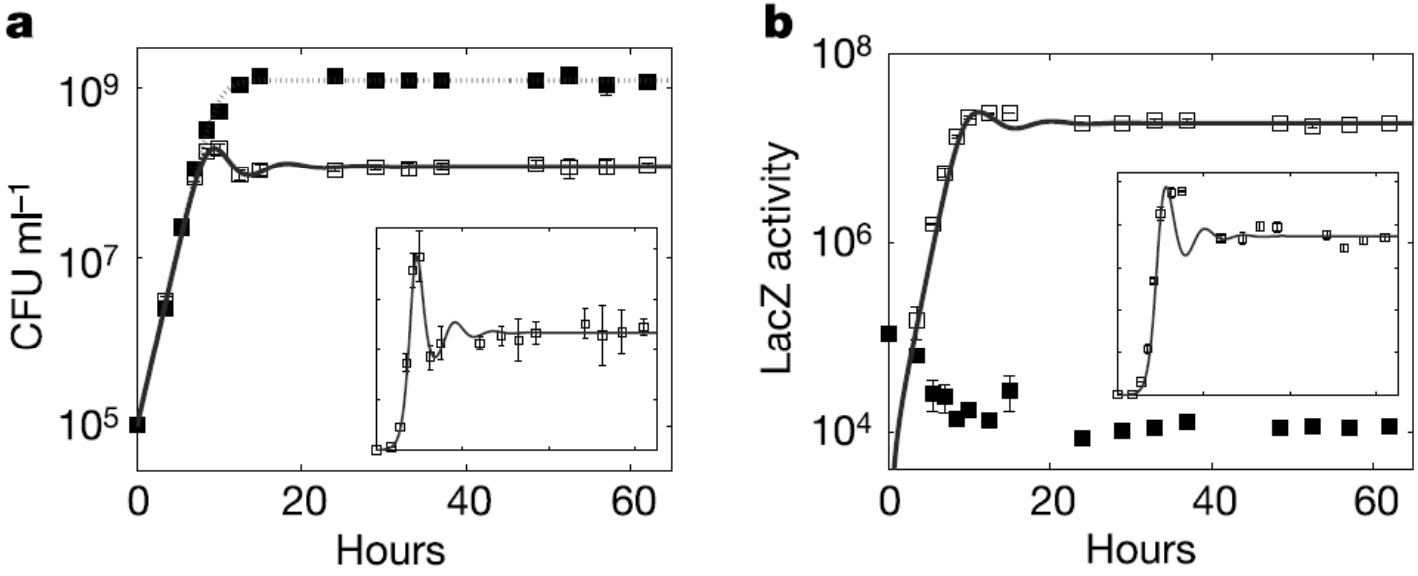


Figure S2:

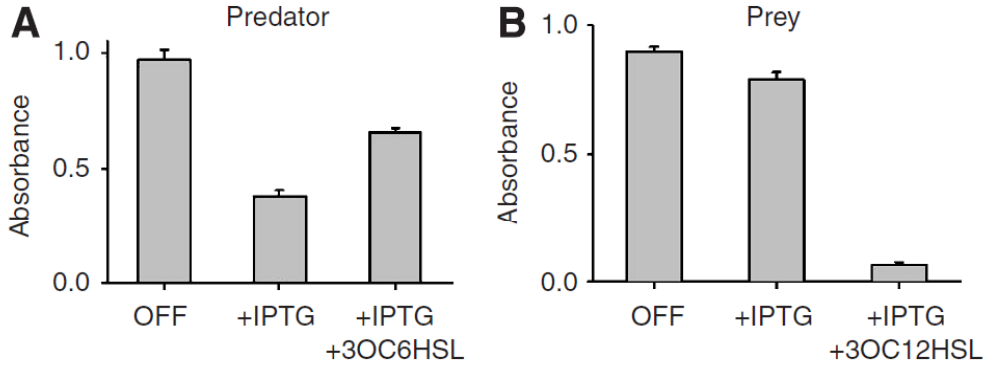
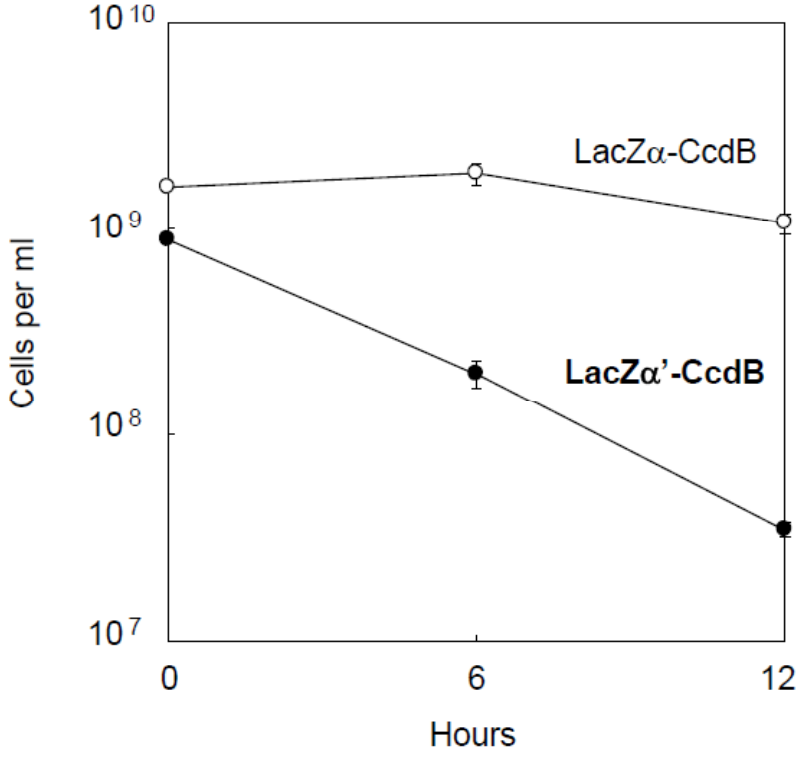


Figure 1 Individual growth behaviors (without interactions) of (**A**) predator and (**B**) prey cells in liquid media. For each condition, 6 ml LBK medium containing chloramphenicol and kanamycin was inoculated with a single bacterial colony and was divided into three 2 ml cultures: 'OFF' cultures contained no inducers, '+IPTG' cultures contained 1 mM IPTG and '+IPTG+AHL' contained 1 mM IPTG and 100 nM AHL, respectively. After 20 h of incubation (light gray bars), optical densities (ODs) of these cultures were measured with a microplate reader (see Supplementary information). Error bars represent standard deviation of triplicate cultures.



Wiki

- Team project description
- Notebook
  - Meetings
  - Lab Work
  - Sample Naming Sheets
- Other
  - Team
  - Project
  - Parts
  - Modeling
  - Human Practice

Calendar of Events

IGEM 2009 Calendar of Events

Note: Dates in grey have not been finalized yet. Make sure to check the calendar periodically for any changes!

19 February	IGEM 2009 registration opens
31 March	IGEM 2008 registration closes; Team registration fee due
13 May	DNA Distribution sent to teams **(target deadline; *subject to change*)**
16/17 May	iGEM Workshop, MIT, USA
1 June	Visa invitation letter requests due
20/21 June	iGEM Workshop, Europe
27/28 June	iGEM Workshop, Asia
15 June	Preliminary team rosters due
1 August	Team project descriptions due
TBD	Jamboree attendance fee due
TBD	Request for variance due (notice and description of any use of non-standard parts or devices schemes due
	Track selection due
	Project abstracts due
TBD	Team rosters due
TBD	Project and part documentation due, including documentation for all medal criteria
	BioBrick Part DNA needs to be received by the Registry
30 Oct - 2 Nov	iGEM Competition Jamboree, MIT, USA

Team:USTC/Notebook

Contents [hide]

Meetings

All the reports information, slides and audio records of our meetings are arranged here.

December

M	T	W	T	F	S	S
1	2	3	4	5	6	7
8	9	10	11	12	13	14
15	16	17	18	19	20	21
22	23	24	25	26	27	28
29	30	31				

January

M	T	W	T	F	S	S
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12	13	14	15	16	17	18
19	20	21	22	23	24	25
26	27	28	29	30	31	

February

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23	24	25	26	27	28	

March

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23	24	25	26	27	28	29
30	31					

April

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20	21	22	23	24	25	26
27	28	29	30			

May

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				1	2	3
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18	19	20	21	22	23	24
25	26	27	28	29	30	31

June

M	T	W	T	F	S	S
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8	9	10	11	12	13	14
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29	30					

July

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		1	2	3	4	5
6	7	8	9	10	11	12
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20	21	22	23	24	25	26
27	28	29	30	31		

August

M	T	W	T	F	S	S
					1	2
3	4	5	6	7	8	9
10	11	12	13	14	15	16
17	18	19	20	21	22	23
24	25	26	27	28	29	30
31						

September

M	T	W	T	F	S	S
	1	2	3	4	5	6
7	8	9	10	11	12	13
14	15	16	17	18	19	20
21	22	23	24	25	26	27
28	29	30				

October

M	T	W	T	F	S	S
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19	20	21	22	23	24	25
26	27	28	29	30	31	

June							July							August							September							October						
M	T	W	T	F	S	S	M	T	W	T	F	S	S	M	T	W	T	F	S	S	M	T	W	T	F	S	S	M	T	W	T	F	S	S
1	2	3	4	5	6	7			1	2	3	4	5					1	2		1	2	3	4	5	6				1	2	3	4
8	9	10	11	12	13	14	6	7	8	9	10	11	12	3	4	5	6	7	8	9	7	8	9	10	11	12	13	5	6	7	8	9	10	11
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29	30						27	28	29	30	31			24	25	26	27	28	29	30	28	29	30					26	27	28	29	30	31	
														31																				

Brainstorming

- 2009-02-10
- 2009-03-07
- 2009-03-22
- 2009-03-30

Lab Work

We chose to use iPad as our Electronic Lab Notebook. iPad is recommended on OWW [1].

Our work progress is also updated here.

May							June							July							August							September							October						
M	T	W	T	F	S	S	M	T	W	T	F	S	S	M	T	W	T	F	S	S	M	T	W	T	F	S	S	M	T	W	T	F	S	S	M	T	W	T	F	S	S
				1	2	3	1	2	3	4	5	6	7			1	2	3	4	5						1	2		1	2	3	4	5	6				1	2	3	4
4	5	6	7	8	9	10	8	9	10	11	12	13	14	6	7	8	9	10	11	12	3	4	5	6	7	8	9	7	8	9	10	11	12	13	5	6	7	8	9	10	11
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25	26	27	28	29	30	31	29	30						27	28	29	30	31			24	25	26	27	28	29	30	28	29	30						26	27	28	29	30	31	
																					31																					

Sample Naming Sheets

We chose to use this wiki page to manage samples. The names are generated with a program written by Jian Zhan.

Instructional Videos

Workshop videos: Videos from the iGEM 2009 spring workshop @ MIT are available below for streaming and download, and higher quality downloads will follow shortly. You can also visit our iGEM channel @ *blip.tv*.

Welcome to IGEM

low ~~high~~

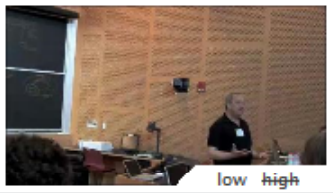
A short welcome and introduction to iGEM by Randy Rettberg

Navigating the Registry

low ~~high~~

Reshma and Randy discuss navigating through partsregistry.org

Introduction to Synthetic Biology



low ~~high~~

Tom Knight gives an introduction to parts based synthetic biology

Project Ideas

slides low ~~high~~

Reshma Shetty gives suggestions on how teams may want to come up with project ideas

Changes for iGEM 2009

low ~~high~~

Randy Rettberg discusses the changes that have taken place for iGEM 2009, the requirements for the teams, and judging the competition

Standard Assembly

slides low ~~high~~

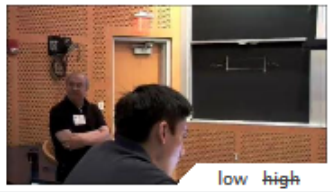
Reshma Shetty shows how parts on the registry are designed for standard assembly

Promoters

low ~~high~~

Barry Canton discusses the promoter category of parts in the registry

Making and Adding Parts



low ~~high~~

Reshma and Meagan show how to make and add parts to the registry

Devices

Barry Canton discusses devices in the registry

Favorites and Shipping Parts

Meagan Lizarazo shows how to make your parts "Favorites" and ship them

low ~~high~~

using the registry

low ~~high~~

Measurements

low ~~high~~

Barry Canton discusses the importance of measuring and documenting the parts on the registry

2009 Distribution, QC, and Sequencing

slides low ~~high~~

Paul and Vinoo discuss an overview of the creation of the 2009 distribution and the quality control process. Randy discusses the sequencing tools on the registry.

Software Tools Track

low ~~high~~

Randy Rettberg discusses the software track for iGEM participants, as well as how software tools are integrated into the registry

Safety in iGEM

screen ipod

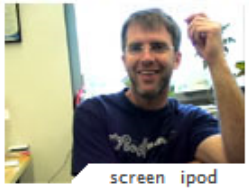
"What safety precautions should my team be taking while participating in iGEM? Why is this important?"

Drew Endy: Defining Synthetic Biology

screen ipod

"Make it easier to build things. Define the things you are building with by using standards. Hide biological complexity with abstraction."

Drew Endy: Believe in Synthetic Biology



screen ipod

"Why should you consider changing how you engineer biological systems from doing ad-hoc research to something that's a more scalable engineering framework?"

Drew Endy: What is a Standard Biological Part?

"What does it mean to have a Standard Biological Part – for example, a BioBrick-standard

Episodes Archive

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Syndicate Show

iGEM 2009 Spring Workshop: MIT Tom's talk on parts based synthetic biology

iGEM 2009 Spring Workshop: MIT Randy's Welcome Speech

Safety in iGEM

Jam07 - Beginnings and Beyond

Jam07 - From Challenge to Triumph

Interview with Alja Oblak from the iGEM06 Ljubljana Team

Brown iGEM07 Team - Introduction to iGEM

Jam07 - What can synthetic biology do for you?

Jam07 - Calgary - "Developing A Genetic Printer"

Jam07 - Caltech - "Selection for Infection"

iGEM 2007 Jamboree Good Times

iGEM 2007 Jamboree Dance Off

Episodes Archive

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Syndicate Show

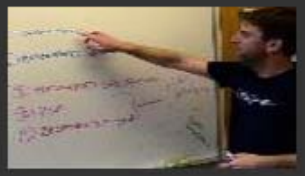
Jam07 - ETH Zurich - "educatETH E.coli System"

tutorial 4.3 - Reviewing your part

Tutorial 4.2 - Entering Part Sequence and Features

Tutorial 4.1 - Adding and documenting a basic part

iGEM Explainer 03 - Drew Endy: What is a Standard Biological Part



iGEM Explainer 01 - Drew Endy: Defining Synthetic Biology

iGEM Explainer 02 - Drew Endy: Believe in Synthetic Biology

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Thank You